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A microanatomical and histological study of the scales of the Devonian sarcopterygian *Miguashaia bureaui* and the evolution of the squamation in coelacanths

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ABSTRACT

Coelacanths have traditionally been described as morphologically_{ly} conservative throughout their long evolutionary history, ~~which~~ ~~that~~ spans more than 400 million years. After an initial burst during the Devonian, a morphological stasis was long thought to have prevailed since the Carboniferous, as shown by the extant *Latimeria*. New fossil discoveries have challenged this view, with punctual and sometimes unusual departures from the general coelacanth Bauplan. The dermal skeleton is considered to represent one, if not the main, example of morphological stasis in coelacanth evolution and as a consequence, has remained poorly surveyed. The ~~flagrant~~ lack of paleohistological data on the dermoskeleton has resulted in a poor understanding of the early establishment and evolution of the coelacanth squamation. Here we describe the scales of *Miguashaia bureau* from the Upper Devonian of Miguasha, Québec (Canada), revealing histological data for a Palaeozoic coelacanth in great detail ~~and for the first time~~, and adding to our knowledge on the dermal skeleton of sarcopterygians. *Miguashaia* displays rounded scales ornamented by tubercles and narrow ridges made of dentine and capped with enamel. At least two generations of superimposed odontodes occur, which is reminiscent of the primitive condition of stem osteichthyans like *Andreolepis* or *Lophosteus*, and onychodonts

like *Selenodus*. The middle vascular layer is well developed and shows traces of osteonal remodelling. The basal plate consists of a fully mineralized lamellar bone with a repetitive rotation pattern every five layers indicating a twisted plywood-like arrangement of the collagen plies. Comparisons with the extant *Latimeria* and other extinct taxa show that these features are consistently conserved across coelacanth evolution with only minute changes in certain taxa. The morphological and histological features displayed in the scales of *Miguashaia* enable us to draw a comprehensive picture of the onset of the coelacanth squamation and to propose and discuss evolutionary scenarios for the coelacanth dermoskeleton.

Keywords: coelacanth, dermoskeleton, histology, odontodes, Palaeozoic, scales

INTRODUCTION

Coelacanths (or actinistians) have frequently been described as displaying a remarkable morphological conservatism following an early evolutionary burst at their emergence in the fossil record at the beginning of the Devonian period (around 420 million years ago) (Forey, 1984; Cloutier, 1991a; Zhu et al., 2012). The discovery in 1938 of *Latimeria chalumnae* (Smith, 1939) off the coast of South Africa was completed by the identification of a second species, *L. menadoensis* (Erdmann et al., 1998; Pouyaud et al., 1999) from the Sulawesi Island of Indonesia, allowing access to extant material for key morphological comparisons with the relatively rich fossil record of coelacanths (Cloutier & Forey, 1991; Forey, 1998). Genomic data obtained from these extant representatives suggest that there is indeed low molecular diversity within populations of *L. chalumnae* (Lampert et al., 2012) and low substitution rates in the coelacanth lineage that led to the two extant species (Amemiya et al., 2010, 2013; Lang et al., 2010; Higasa et al., 2012). Molecular data have also revealed the existence of a putative third coelacanth species in Indonesia (Kadarusman et al., 2020). However, the supposed morphological stasis of coelacanths has been challenged by new fossil discoveries (Casane & Laurenti, 2013) that highlighted important differences in the skull, axial skeleton, and internal organs

throughout their evolutionary history (Lund & Lund, 1984, 1985; Forey, 1998; Friedman & Coates, 2006; Brito et al., 2010; Wendruff & Wilson, 2012; Cavin et al., 2017; Cupello et al., 2017).

In the case of the dermal skeleton, the alleged conservatism of coelacanths has pervaded anatomical descriptions and led to a flagrant lack of comparative histological information on fossil coelacanth scales, with Forey (1998, p.220) even stating that 'there is little to suggest that they vary in any significant way from the structure seen in *Latimeria*'. Indeed, the scales of all known extinct and extant actinistians are rounded and are ornamented with tubercles and/or coarse or thin ridges that may break into small leaf- or spoon-shaped tubercles near the anterior margin of the exposed area in early forms. This ornamentation pattern is uniformly maintained in many coelacanths from the Palaeozoic to Recent (e.g. *Gavinia*, *Miguashaia*, *Diplocercides*, *Nesides*, *Rhabdoderma*, *Whiteia*, *Piveteauia*, *Undina*, *Macropoma*, *Coelacanthus*, *Swenzia*, and *Latimeria*) (Stensiö, 1937; Schaeffer, 1941; Forey & Young, 1985; Cloutier, 1996; Forey, 1998; Forey et al., 2000; Clément, 1999, 2005; Long, 1999) (Table 1).

The histological nature of the ridges and tubercles of the scales and dermal bones of coelacanths remains unclear in many taxa and might lack the seeming homogeneity implied by external observation. In the extant *Latimeria*, the ridges and

tubercles have been extensively surveyed and illustrated (Millot & Anthony, 1958; Millot et al., 1978) and are of odontogenic nature, as they are made of dentine capped with enamel (Bernhauser, 1961; Smith et al., 1972; Castanet et al., 1975; Meunier et al., 2008; Meunier et al., 2019). In extinct forms, the ridges have been reported to be made of dentine in *Diplocercides jaeckeli* from the Devonian (Gross, 1935, 1966), *Undina penicillata* from the Jurassic (Gross, 1935, 1966) and *Macropoma lewesiensis* from the Cretaceous (Williamson, 1849). In *Diplurus newarki* from the Triassic (Schaeffer, 1952) the scales were tentatively described as being covered by a thin layer of enamel, except on the ridges and tubercles. In ~~the dermal bones of~~ *Spermotodus pustulosus* from the Permian (Westoll, 1939), the dermal bones display the tubercles ~~are~~ composed of dentine and enamel ~~(i.e., odontodes)~~ (Meinke, 1982) organized in displaying several layers (i.e. of superimposed odontodes) with t-and traces of resorption and redeposition of bone and odontodes, but the scales were not surveyed. Despite this wealth of data and material availability, no comprehensive and comparative histological work has been done on the scales of coelacanths and thus many observations, not properly illustrated by histological preparations, might be considered as unreliable in the absence of a thorough histological survey. Most notably, Gross (1935) already pointed out that the histology of coelacanths had been poorly surveyed and comparative data from Palaeozoic, and

especially Devonian coelacanths were crucially lacking for comparisons with more recent taxa.

The Devonian fossil record of coelacanths is scarce, represented by taxa such as *Eoactinistia foreyi* from the Pragian of Australia (Johanson et al., 2006), *Euporosteus* from the Pragian-Givetian of China (*E. yunnanensis*) and Germany (*E. eifeliensis*) (Stensiö, 1937; Zhu et al., 2012), *Gavinia syntrips* from the Givetian of Australia (Long, 1999), *Holopterygius nudus* from the Givetian of Germany (Jessen, 1973; Friedman & Coates, 2006), *Diplocercides* from the Frasnian of Germany (*D. kayseri*, *D. heiligenstockiensis*, *D. jaeckeli*) and the Visean (Carboniferous) of Scotland (*D. davisii*) (Stensiö, 1922, 1937; Moy-Thomas, 1937; Jessen, 1966; Forey, 1981), *Miguashaia* from the Frasnian of Québec (Canada) (*M. bureauui*) and Latvia (*M. grossi*) (Schultze, 1973; Cloutier, 1996; Forey et al., 2000), *Chagrinia enodis* and *Coelacanthus welleri* from the Famennian of eastern USA (Eastman, 1908; Schaeffer, 1962), and *Serenichthys kowiensis* from the Famennian of South Africa (Gess & Coates, 2015). Early coelacanth fossil material is not only rare but also generally poorly preserved, compressed, or simply too precious to allow histological sectioning. Abundantly preserved individuals displaying a good histological quality are even scarcer, dramatically reducing the access to paleohistological data. One of the few exceptions to this limited source of information is the material from Miguasha (Québec, Canada).

The Miguasha fossil site has yielded exquisitely preserved fossil specimens from a wide range of early vertebrates, including the iconic *Eusthenopteron foordi* (Jarvik, 1944, 1954, 1980) and *Elpistostege watsoni* (Westoll, 1938; Cloutier et al., 2020). *Miguashaia bureau* (Schultze, 1973; Cloutier, 1991a,b, 1996) represents the only coelacanth found in Miguasha so far. It was a main predator in the estuarine paleoenvironment of the Escuminac Formation (Chevrinais et al., 2017a), which is dated as middle Frasnian in age (Cloutier et al., 1996). The vertebrate specimens from Miguasha are numerous and frequently three-dimensionally preserved, which has given an excellent account of their external morphology (Cloutier et al., 2011, Cloutier, 2013). Likewise, histological features are represented either as hard tissues (e.g. enamel, dentine, cellular bone, cartilage) or as cell spaces (e.g. osteocyte and chondrocyte lacunae), sufficiently well-preserved to allow genome size inference (Thomson, 1972; Organ et al., 2016), as well as soft tissues preservation (Cloutier, 2013). Previous microstructural and histological studies have yielded excellent results on several taxa from the Escuminac Formation, such as the anapsid *Euphanerops longaevus* (Janvier & Arsenault, 2002; Chevrinais et al., 2018), the osteostracan *Escuminaspis laticeps* (Janvier et al., 2004), the 'placoderm' *Bothriolepis canadensis* (Burrow, 2005; Downs & Donoghue, 2009), the 'acanthodian' *Triazeugacanthus affinis* (Chevrinais et al., 2017b), the

sarcopterygians *Eusthenopteron foordi* (Schultze, 1969; Laurin et al., 2007; Zylberberg et al., 2010; Meunier & Laurin, 2012) and *Scaumenacia curta* (Thomson, 1972; Smith et al., 1987), and the actinopterygian *Cheirolepis canadensis* (Zylberberg et al., 2015; Meunier et al., 2018).

Here, we study the ~~histological-morphological, and~~ microanatomical, ~~and histological~~ organization of the scales of the early coelacanth *Miguashaia bureau* from the middle Frasnian of eastern Québec (Canada). It constitutes ~~a the~~ ~~first~~ thorough investigation of the dermal scales of a Palaeozoic coelacanth and provides much needed comparative information on the early establishment of the histological features of the actinistian squamation. Comparison with other extinct coelacanths and with the extant *Latimeria* will provide a more complete picture of the histological diversity in actinistians and better document the evolution of the dermal skeleton in sarcopterygians and early osteichthyans. With these detailed descriptions and holistic approach, we hope to invigorate the study of the paleohistological diversity of the dermal skeleton of actinistians, which has been largely overlooked in most analyses of coelacanth evolution.

MATERIAL AND METHODS

Material

Miguashaia

Original and newly discovered material of *Miguashaia bureaui* has been studied, focusing on the external morphology of the scales: MHNM 06-41ab, MHNM 06-494 (mean scale diameter: 1.87 cm; total length of specimen: 37.5 cm), MHNM 06-641, MHNM 06-1232, MHNM 06-1233, MHNM 06-1234, MHNM 06-1236ab, MHNM 06-1237; MHNM 06-1239; MHNM 06-1318ab; MHNM 06-1633 (mean scale diameter: 0.16 cm; total length of specimen: 6.98 cm), MHNM 06-1809ab (mean scale diameter: 1.28 cm; total length of specimen: ca. 43 cm); MHNM 06-2414, ULQ 120ab (holotype). Fossil material used for the histological investigation consists of two separated blocks (MHNM 06-1238; MHNM 06-1495) containing numerous articulated trunk scales. Specimen MHNM 06-1238 (Fig. 1A) displays disarticulated axial and appendicular elements including at least two neural arches (n.ar) and probably haemal spines, caudal fin radials (ra), and a few fine lepidotrichia (lp) most likely belonging to the second dorsal fin. Specimen MHNM 06-1495 (Fig. 1B) shows disarticulated fragments of the opercular series (op.s) and pectoral girdle elements (cleithrum, cl), most of them preserved as imprint_s of the smooth inner surface of the dermal bones, and some segments of pectoral lepidotrichia (lp) with the characteristic interlocking system. Scales (sc) of specimen MHNM 06-1495 are most likely from the anterior part of the trunk as well as some anteroventral scales. Scales (sc) of specimen MHNM 06-1238 are most likely from the posterior part of the flanks, posteroventral to the second dorsal fin.

All specimens come from the middle Frasnian Escuminac Formation, Miguasha, Québec, Canada. All the fossil material is housed at the Musée d'Histoire naturelle de Miguasha (MHNM), parc national de Miguasha, Québec (Canada) with the exception of the holotype that is housed at the Université Laval (ULQ), Québec City, Québec (Canada).

Latimeria

Latimeria chalumnae. The morphology (SEM) and the microstructure (ground sections) of the scales have been examined predominantly from two main specimens: (a) the scales overlying the basis of the pelvic fin of MNHN-ZA-AC-2012-26 (CCC79, adult female, 78 kg, 163 cm TL) and (b) several scattered scales of specimen MNHN-ZA-AC-2012-29 (CCC117, adult female, 19 kg, 109,5 cm TL), both captured off the Comoro Islands (Nulens et al. 2011). The scales were fixed in neutral formalin and kept in 70° ethanol. All specimens are housed at the Muséum national d'Histoire naturelle (MNHN) of Paris (France).

Latimeria menadoensis. Nine scales were sampled from the second known Indonesian extant coelacanth specimen (MZB10003) (CCC 175; adult female 29,2 kg, 124 cm TL) captured off Manado Tua, Sulawesi Island (Nulens et al, 2011). The scales were originally fixed in formalin, and then stored in 70° ethanol. After measurements of the odontodes, some scales were studied with SEM and others were used for ground sections. The

specimen is housed at the Museum Zoologicum Bogoriense (MZB) (Indonesian National Zoological Museum) in Bogor (Indonesia).

Institutional abbreviations: **FMNH**, Field Museum of Natural History (Chicago, USA); **MHNM**, Musée d'Histoire naturelle de Miguasha (Miguasha, Canada); **MNHN**, Muséum national d'Histoire naturelle (Paris, France); **MZB**, Museum Zoologicum Bogoriense (Bogor, Indonesia); **ULQ**, Université Laval (Québec City, Canada); **YPM**, Peabody Museum, Yale University (New Haven, USA).

Methods

Miguashaia

Ground sections were performed on both fossil specimens (MHNM 06-1238 and MHNM 06-1495) at the MNHN. The fossil material was embedded in polyester resin (GBS 1; Brot) and sectioned for study. The various sections were ground and polished to a thickness of 60–80 µm and observed under transmitted natural light with an Olympus BX51 microscope. Pictures were taken with a digital camera Olympus Camedia C-5060 and finalized in Adobe Photoshop.

Latimeria

For the SEM observations, the scales were steeped in 6 or 12% sodium hypochloride solution at room temperature (in order to strip the surface of the scales and to destroy the

unmineralized collagenous fibres, thus cleaning the mineralized front of the basal plate). The scales were then washed in distilled water, dehydrated in absolute ethanol, air-dried, and coated with evaporated gold. The external and internal surfaces of the scales were examined in a JEOL-SEM-35 scanning electron microscope. Thin sections of the scales (30 to 100 μm in thickness) were observed in polarized light with a Zeiss Axiovert 35 microscope. Microradiographies were performed on non-decalcified sections (focal point-object distance 10 cm, power 10 to 15 Kv at 7 mA). Histological sections after decalcification with EDTA (ethylenediaminetetraacetic acid) and inclusion in paraffin (sections 5 to 10 μm in thickness) were treated with the following stains: hemalun-picro-indigocarmin, APS, toluidine blue, Prenant's trichrome.

DESCRIPTION

Morphology

The external and internal surfaces of the scales match reasonably the previous description by Cloutier (1996) (Fig. 2). All scales of *Miguashaia bureaui* are almost rounded, slightly broader than long, and both sides of the scales at mid-length are linear and parallel.

The ornamented area (posterior field) occupies approximately two fifths of the area of the scale; it is

approximately twice as broad as long. The ornamentation pattern of *M. bureau*i varies according to the position of the scales on the body as well as during ontogeny (Fig. 2). Five types of ornamentation could be observed on the outer ornamented field: (1) coarse tubercles (co.tu), (2) coarse and short longitudinal ridges (co.ri), (3) fine leaf- or spoon-shaped tubercles (sp.tu), (4) fine, parallel, longitudinal, low-profile ridges (f.ri), and (5) smooth, thinly and regularly pitted enamel surface (pi.e).

The coarse tubercles (co.tu) vary in shape from circular to ovoid. Occasional coarse tubercles located near the anterior-most limit of their distribution share the leaf-shape of the fine tubercles (Fig. 2C). The coarse tubercles are also found on the pectoral girdle and lateral gulars of *M. bureau*i. Although we consider that the coarse tubercles are a distinct category of ornamentation from the coarse ridges, these two types of ornamentation form a continuum. The coarse and short longitudinal ridges share similarities with the coarse tubercles. The coarse ridges (co.ri) are either linear or slightly curved, with the posterior extremity forming an apex. No bifurcation has been observed with the coarse ridges. In contrast to the fine low-profile ridges, the space between coarse ridges is broader than with the fine ridges. The longest ridges are found in the median region of the ornamented field. The median patch of coarse ridges is surrounded laterally and posteriorly by coarse tubercles. The

coarse ornamentation has an extremely restricted distribution on the body (Fig. 2B-C).

The fine leaf- or spoon-shaped tubercles (sp.tu) occur only at the limit between the overlapped area (anterior field) and the ornamented area (posterior field). The tubercles are lanceolate to ovate and medially grooved with the apex pointing posteriorly. The size of these tubercles varies within a scale (Fig. 2C-E). Similar tubercles located in a similar position are also found in *Miguashaia grossi* (Forey et al., 2000, fig. 6B: most likely the two insets of figure 6B have been inverted).

The fine ridges (fi.r) are mostly linear and without bifurcation (Fig. 2D1,E). There are five (e.g. MHN 06-2414) to six (e.g. MHN 06-1233) ridges per mm. Individual ridges are approximately 0,16 mm in width. Most ridges are linear; however, marginal ridges may be slightly curvilinear in the largest specimens. On each scale, there are few ridges that bifurcate anteriorly or posteriorly. Occasionally, adjacent ridges merge along their length, thus keeping their respective width. Along the merged sections, spaces are left unmerged forming small pores (0.16-0.18 mm in length and 0.03 mm in width) that result in a pitted surface. In contrast to *M. bureaui*, the longitudinal ridges of *M. grossi* radiate from the medial axis of the scale (Forey et al., 2000, fig. 6B).

The pitted enamel surface (pi.e) covers from the posterior margin of the posterior field up to the complete ornamented

area (Fig. 2E). The pores are ovoid to fusiform; the origin of the pores comes from the merging of adjacent ridges. The pores are aligned longitudinally in the prolongation of the inter-ridge space as well as across the width of the scale. Along a longitudinal alignment, there is a spacing of approximately 0.25-0.27 mm. During growth, this condition originates at the posterior margin of the scales and progresses anteriorly up to the condition in which the complete ornamented area is covered by a pitted surface. The pitted enamel surface was referred as the "almost continuous enamel covering" by Forey (1998, p. 329).

In agreement with Cloutier (1996), the ornamentation of the posterior field of a scale differs in relation with its position on the body. Few scales from the anteroventral region of the body bear coarse tubercles and coarse ridges (MHNM 06-41, MHNM 06-641, MHNM 06-1809) (Fig. 2C). Flank scales from the anterior region of the trunk present a narrow area of small leaf- or spoon-shaped tubercles (anteriorly) and fine longitudinal ridges and a pitted enamel surface (posteriorly). Flank scales from the posterior region of the trunk are either covered by (1) fine leaf- or spoon-shaped tubercles and fine longitudinal ridges (Fig. 2D1) (MHNM 06-1633, MHNM 06-1236), (2) fine leaf- or spoon-shaped tubercles, fine longitudinal ridges and pitted enamel surface (MHNM 06-1234), (3) fine longitudinal ridges and pitted enamel surface (Fig. 2E) (MHNM 06-494, MHNM 06-1265, MHNM 06-1809), or (4) solely pitted

enamel surface (MHNM 06-1237, MHNM 06-1265). The extent of the pitted enamel surface increases during growth in order to cover the complete ornamented area. Smaller scales from the caudal lobe only bear fine longitudinal ridges in both small and large individuals (MHNM 06-1318, MHNM 06-1809). Although the ornamentation is not preserved on the complete specimens, the smallest individuals appear to only have fine longitudinal ridges (ULQ 120, MHNM 06-1633). The coarse tubercles and ridges seem to develop only in larger individuals (MHNM 06-641).

The inner surface of the scales is flat and characterized by small circular bumps (ci.b) perforated in their centre (Fig. 2D2); these structures had already been reported and illustrated by Cloutier (1996, fig. 17D). Although small, these bumps vary slightly in diameter. They do not seem to be aligned. These bumps are located at the level of the outer ornamented area (posterior field). As far as we know, *Miguashaia bureaui* is the only sarcopterygian species with this feature. The posterior part of the inner surface shows fine concentric *circuli* (growth lines) with some variations in terms of width bands and inter-band spacings.

Sensory lateral line scales had been reported by Schultze (1973) and Cloutier (1996) on a juvenile specimen (holotype ULQ 120). While Schultze (1973, fig. 1a) illustrated scales with one central elongated canaliculus per scale forming a horizontal trajectory starting at the posteroventral limit of

the anocleithrum, Cloutier (1996, fig. 1a) illustrated scales with one central elongated canaliculus associated with two pores above and below forming also a horizontal trajectory on most of the length of the trunk but slightly curving dorsally along the anocleithrum. Seven to eight scales belonging to the anterior section of the main lateral line canal are preserved on specimen MHNM 06-1236 starting above the dorsal part of anocleithrum. Each scale has the main elongated canaliculus flanked dorsally and ventrally by three or four slightly curving canals.

Histology

In both specimens used for the histology study (MHNM 06-1238 and MHNM 06-1495) (Fig. 1), the scales are imbricate and preserved in articulation. The inner surface of scales is best preserved in specimen MHNM 06-1238 showing the complete shape of the scales (average diameter of 1.5 cm) (sc, Fig. 1A). The inner surface is generally smooth and displays a few weakly marked growth rings and scattered bumps in the central and posterior regions (Fig. 2D2). In addition, there is a patch of imbricated scales showing the outer field with the fine longitudinal ridges. The scales from MHNM 06-1495 (average diameter of 1.7 cm) are visible on their external surface (sc, Fig. 1B). Most scales are preserved as imprints showing either fine longitudinal ridges or pitted enamel surface; these scales are most likely from the anterior region of the flanks.

A few well-preserved scales (most likely anteroventral scales) show large tubercles as well as leaf-shaped tubercles and fine longitudinal ridges.

The exposed area (posterior field)

The scales can be microstructurally divided into three well defined superimposed portions (Fig. 3): 1) a superficial layer of odontodes (od) made of enamel (e) and dentine (d) (Figs 3A, 4A-C), 2) a middle layer (ml) of vascular, woven-fibered bone (vb) with numerous vascular canals (i.e., spongiosa) (Figs 3B, 5, 6), and 3) a basal plate (bp) made of parallel-fibered lamellar bone (lb) (i.e., isopedine) (Figs 3C, 7). The enamel and dentine layers form the tubercles and the ridges ornamenting the exposed area. Certain odontodes display a pointed or concave lenticular outline in cross-section (Figs 3A, 4B-E).

Enamel. The enamel is a single uniformly developed sheet of approximately 15-25 μm in thickness (e, Figs 3A, 4B-E). No cytoplasmic prolongations from the dentine are present in the enamel, thus ruling out the occurrence of enameloid. The enamel only occurs in the tubercles and is restricted to their tips. No continuous sheet of enamel has been distinctly observed in our two sectioned specimens but we suspect that it displays identical features to the enamel of the odontodes.

Dentine. The dentine layer forms the base of the odontodes (d, Figs 3,4) and is variable in thickness (between 50-100 μm).

Dentinal tubules (oc, odontoblastic canaliculi) (Fig. 4B) are roughly perpendicular to the enamel layer; these radiate from small odontoblastic cavities (pc, pulp cavities) located at the base of the dentine layer (Fig. 3A). The dentine of *Miguashaia bureaui* can thus be described as orthodentine (Francillon-Vieillot et al., 1990). At least two generations of superimposed odontodes occur in some sections (Fig. 3A, 4D-E), with partial resorption of the older generation by the surrounding cellular bone. Cosmine is confidently considered absent as there is no evidence of a pore-canal system developed between the enamel, dentine and vascular bone.

Spongiosa. A middle vascular bone layer (i.e., spongiosa) occurs between the odontodes of the superficial layer and the isopedine layer of the basal plate. It is formed of woven-fibered bone pervaded by numerous primary vascular canals (vb, Figs 3A-B, 4B-E, 5). As the dentine layer, the spongiosa is well-developed but varies in thickness (approximately between 50-100 μm) across the scales. Numerous osteocytic lacunae with their ramified canaliculi can be seen (Figs 3B, 4D, 6D-F). Horizontally-connected vascular canals are located below the dentine and surround the buried odontodes. Some of these vascular canals open through the external surface (vc, Fig. 4B-C). The woven-fibered bone of the spongiosa forms the radial striae of the overlapped area of the scales (vb, Fig. 6A) in the form of pointed or mushroom-like projections of compact bone (vb, Figs 5B, 7A). Secondary vascular canals can

be seen (Fig. 6C,D,E) indicating the presence of localized osteonal remodelling.

Isopedine. The basal plate is made of a fully mineralized, lamellar bone with a plywood-like structure with a variable orientation of the collagen plies (1b, Figs 3C, 4C-E, 7B). The isopedine layer accounts for one third to half of the scale total thickness (approximately between 100-150 μm). The plies of the basal plate are made of thick collagenous fibres, organized in bundles that appear roughly quadrangular in cross section (Fig. 3C). There is only one layer of fibres per ply with each ply being about 10 to 20 μm thick. Horizontal sections of the basal plate confirm that collagenous fibres are parallel to each other in a given ply. The direction of the fibres rotates from a given ply to the next (Figs 3C, 7B). We have observed the presence of five fibrillary directions (Fig. 8A), indicating that a repetitive pattern occurs every five layers (Figs 3C, 7B) and that each fibre rotates from one layer to the next with an angle of approximately 36° , thus constituting a twisted plywood. But contrary to the basal plate of the scales of *Latimeria* (Fig. 8B,C), no arciform patterns, characteristic of a double-twisted plywood (Giraud et al., 1978a,b; Meunier, 1981; Mondéjar Fernández & Meunier, 2020), occur in transversal section, thus indicating that the isopedine of the scales in *Miguashaia bureaui* is composed of a simple twisted plywood. Several small osteocytic lacunae and canaliculi are visible between the plies, showing that the

collagenous network enclosed elongate osteocytes (elasmocytes *sensu* Meunier, 1984; Francillon-Vieillot et al., 1990) with their cytoplasmic processes extending along the collagen fibres (Figs 4C-D, 8A). Many vertical vascular canals orthogonally cross the isopedine layer and several open through the internal surface forming a bulge (Fig. 7B), corresponding to the perforate small circular bumps of the internal surface (Fig. 2C). In some sections, the irregular contact of the collagen plies at the internal (or visceral) margin of the scales (Figs 4A, 5A, 7B) may indicate the occurrence of Mandl's corpuscles attesting to a possible delay between collagenous synthesis and its mineralization (see Schönborner et al., 1981 for teleostean elasmoid scales) in successive mineralization events across the basal layer (Fig. 4A).

The overlapped area (anterior field)

The upper part of the superficial layer (sl) of the area overlapped by the adjoining scales is generally deprived of odontodes, yet it shows a superficial ornamentation composed of a layer of compact acellular bone (Fig. 5A,C) that recalls the external layer of the teleostean elasmoid scales (Francillon-Vieillot et al., 1990; Zylberberg et al., 1992). This external acellular ornamented layer overlays a vascularised layer of cellular bone (Fig. 5B). The vascular canals appear to be more uniformly distributed than in the

exposed area (sl, Fig. 7A) and open regularly onto the surface between the small striae made of woven-fibered bone (Figs 5, 7A). The basal plate has a slightly greater relative thickness than in the posterior field but displays the same organization and a similar number of stacked layers of collagenous fibres (Fig. 5A).

DISCUSSION

Histology of *Miguashaia* and comparisons

Despite their rich fossil record, comprising more than 60 genera and 130 species (Cloutier & Forey, 1991; Forey, 1998), few surveys of the dermal skeleton of coelacanths have been undertaken. Gross (1966:42) first noted that the scales of coelacanths had barely changed their histological and microstructural organisation during their long evolutionary history, a statement that was later reprised in subsequent reviews of coelacanth evolution (e.g. Janvier, 1996; Forey, 1998). To our knowledge, few detailed accounts on the palaeohistology of the scales and dermal bones of extinct coelacanths have been published or properly illustrated. These include most notably histological reports on *Macropoma lewesiensis* (Williamson, 1849), *Coelacanthus granulatus* (Goodrich, 1907), *Undina penicillata* (Gross, 1935, 1966), *Diplurus newarki* and *Chagrinia enodis* (Schaeffer, 1952, 1962), and *Spermatodus pustulosus* (Meinke, 1982). Indeed, many

descriptions of paleohistological features are either solely based on external observation, drawings, or ~~on~~-unfigured thin sections (e.g. Schaeffer, 1952; Forey, 1998). As such, the reliability of many descriptions is difficult to assess due to the lack of proper illustrations ~~since~~because no description can consistently capture all the information contained in a histological section.

The thin, imbricate round scales of all histologically studied coelacanth, as evidenced here in *Miguashaia* (Figs 3, 4A, 7A), display in the posterior field three well-defined portions, differing in their microstructure: 1) a superficial layer of odontogenic components (odontodes generally made of enamel and dentine), 2) a middle layer composed of vascular bone (spongiosa) and 3) a basal plate made of lamellar bone (isopedine). The combination of this microstructural arrangement with a rounded outline allows to ascribe the scales of coelacanth to the elasmoid type (Bertin, 1944; Smith et al., 1972; Castanet et al., 1975; Miller, 1979; Meunier & Zylberberg, 1999; Hadiaty & Rachmatika, 2003; Meunier et al., 2008), a scale morpho- and histotype that is widely spread and convergently acquired in osteichthyans (e.g. Francillon-Vieillot et al., 1990; Schultze, 1977, 2015, 2018; Mondéjar-Fernández & Meunier, 2020). The general structure of the elasmoid scales displayed in *Miguashaia* has been maintained across coelacanth evolution until the Recent as evidenced in *Latimeria* (Castanet et al., 1975; Meunier et al.,

2008), although the ornamentation patterns, ossification rate, and relative thickness of the various layers differ across taxa. Nevertheless, several key differences can be recognized between certain taxa, revealing a significant histological disparity.

The superficial layer comprises the outer ornamentation of the exposed area of the scales, which is composed of pointed to rounded tubercles and/or ridges, variable in width and number within as well as between taxa (e.g. *Miguashaia*, *Diplocercides*, *Spermatodus*) (Table 1). The presence of ridges has been noticed in numerous extinct coelacanths (e.g. *Diplurus*, *Rhabdoderma*, *Axelrodichthys*, *Holophagus*) (Forey, 1998). In many instances, their shiny appearance is suggestive of the odontogenic nature of these ridges, composed either solely of dentine or more likely covered by enamel (e.g. *Diplocercides*, *Rhabdoderma*, *Axelrodichthys*, *Holophagus*). However, few histological studies have been performed ~~on the scales~~ to confirm the putative universal occurrence of dentine, let alone the co-occurrence of enamel (e.g. Gross, 1935). For instance, the scales of *Diplocercides* (Stensiö, 1937) have not been histologically surveyed although Gross (1935:44) stated that in *D. jaeckeli* the small ridges were "certainly made of dentine". The scales of *Macropoma* are ornamented with elongate tubercles, with the largest ones occupying the central position in the exposed area (Forey, 1998, fig. 11.12). Most remarkably, the scales from the

posterior half of the body display large pointed tubercles resembling "hollow cylinders" (Forey, 1998, fig. 11.12B). Williamson (1849, pl. XLIII 27,28) illustrated these tubercles in *M. lewesiensis*, confirming their odontogenic nature. The scales of *Diplurus* (Schaeffer, 1948, 1952) do not possess tubercles, only large ridges, probably odontogenic. Schaeffer (1952:52) stated that in *D. newarki* "the bones, and apparently also the scales, are covered by a thin layer of enamel", but this is difficult to confirm given that thin sections were not illustrated. *Coelacanthus* presents "elongated tubercles or shiny ridges" (Goodrich, 1907:766) that appeared as "hollow" in the poorly illustrated cross sections of *C. granulatus* (Goodrich, 1907, pl.XLIV 12). The odontode nature of these structures was not confirmed, but they might represent dentine tubercles with large pulp cavities capped with enamel or large vascular canals of the superior portion of the spongiosa. The odontodes of *Undina penicillata* (Gross, 1966, fig.7B) are remarkably composed exclusively of dentine and do not present enamel, as opposed to those of *Miguashaia*, *Spermotodus* and *Macropoma*. Noteworthy, *Mawsonia* appears to lack odontogenic ornamentation as the ridged surface of the dermal bones and scales has been described as being solely made of bone in *M. brasiliensis* (Yabumoto, 2002), although histological data have not been published. The ridges and tubercles of *Chagrinia enodis* (Schaeffer, 1962; Cloutier, 1996, fig. 9D) were also described as made of bone, ~~although~~ but according to Forey

(1998), the absence of odontogenic ornamentation might be due to a preservation artefact. Other taxa described as displaying a "rugose" ornamentation of the scales (*sensu* Forey, 1998) such as *Chinlea sorenseni* (Schaeffer, 1967), are not suited to histological survey as the scales are preserved as impressions.

An ornamentation composed of tubercles and/or ridges is thus fairly common and widespread in the scales and dermal bones of coelacanths (Figs 9, 11). However, due to the lack of histological data for many taxa, it is currently difficult to determine whether it is consistently composed of odontogenic tissues and to study its ontogenetic development. In some Mesozoic taxa, the odontodes usually display large but shallow pulp cavities, connected to a horizontal network of vascular canals in the spongiosa (e.g. *Undina*, *Macropoma*). This condition, retained in *Latimeria chalumnae* (Roux, 1942; Ørvig, 1977; Castanet et al., 1975) (Figs 9, 10A), differs from that of *Miguashaia bureau*, in which pulp cavities are reduced (Figs 3A, 4). A possible evolutionary trend can be proposed, from small and numerous odontodes with small pulp cavities in early Palaeozoic coelacanths (e.g. *Miguashaia*) to fewer but larger odontodes with wider pulp cavities in younger taxa from the Mesozoic (e.g. *Undina*, *Macropoma*) to the Recent (*Latimeria*).

Odontodes are rarely found isolated, either on the scales or on the dermal bones. Indeed, superimposed generations of

odontodes embedded in the spongiosa are known in *Miguashaia bureaui* (Figs 3,4), *Spermotodus pustulosus* (Meinke, 1982), and *Undina penicillata* (Gross, 1936, 1966), but not in *Macropoma lewesiensis* (Williamson, 1849). For instance, the presence and distribution of odontodes in the dermal bones of *Spermotodus* is similar to those of *Latimeria*, but in the former the old generations of odontodes are more robustly developed and less resorbed than in the latter (Meinke, 1982). This condition, also present in *Undina*, appears closer to that of early osteichthyans with large partially to non-resorbed odontodes (e.g. *Andreolepis*, *Lophosteus*, *Psarolepis*) (Qu et al., 2013, 2016; Jerve et al., 2016). The occurrence of separated and superimposed odontodes represents the primitive pre-cosmine condition in osteichthyans (Mondéjar-Fernández, 2018) since these odontodes (either in the form of tubercles and/or ridges) are found in stem osteichthyans like *Andreolepis hedei* (Gross, 1968; Qu et al., 2016), *Lophosteus superbus* (Gross, 1969; Jerve et al., 2016), *Ligulalepis toombsi* and *Orvikuina* sp. (Schultze, 1968), *Dialipina* (*D. salgueiroensis* and *D. markae*) (Schultze, 1968, 1977), and *Naxilepis gracilis* (Wang & Dong, 1989). However, the condition of coelacanth may represent a return to the odontode-ornamented condition of certain stem osteichthyans (e.g. *Andreolepis*), probably derived from a cosmine-covered condition in more crownward stem osteichthyans (e.g. *Psarolepis*) and sarcopterygians (e.g. *Styloichthys*, see below), rather than a retention of a pre-

cosmoid state as previously suggested (Smith, 1977, 1979; Schultze, 1977).

The middle layer of vascular bone is variably developed in the scales of coelacanth. In the case of taxa with superimposed odontodes, the older generations are surrounded by vascular bone and may show traces of resorption as seen in *Miguashaia bureau* (Figs 3-6), *Undina penicillata* (Gross, 1966), and *Spermotodus pustulosus* (Meinke, 1982). The vascular bone layer is made of a cellular woven-fibered bone, as evidenced by the occurrence of numerous osteocyte lacunae concentrically arranged around the embedded odontodes and vascular canals (e.g. *Miguashaia*, *Spermotodus*, *Undina*) (Fig. 6C). In *Miguashaia bureau* (Figs 4, 7A), *Macropoma lewesiensis* (Williamson, 1849) and *Undina penicillata* (Gross, 1935, 1966), the vascular canals of the spongiosa open to the surface through pores, and, when present, some of them are connected with the pulp cavities of the dentine. In *Latimeria*, the middle layer is extremely reduced but horizontally-connected vascular canals occur at the base of the odontodes (Smith, 1972; Castanet et al., 1975) (Fig. 10B). In *Undina penicillata*, Gross (1935) described the presence of Sharpey's fibres in the spongiosa, whereas Williamson (1849) described similar structures in *Macropoma lewesiensis* but identified them as dentinal tubules (odontoblastic canaliculi). No distinctive Sharpey's fibres have been confidently identified

in the middle layer nor in the basal plate of the scales of *Miguashaia bureau*.

The basal plate of the scales and dermal bones of actinistians is composed of a layer of lamellar bone with a plywood-like structure (i.e. isopedine). The basal plate is well ossified in the scales of all extinct coelacanths (e.g. *Miguashaia*, *Coelacanthus*, *Undina*, *Macropoma*), but it is not in the extant *Latimeria* (Castanet et al., 1975; Meunier & Zylberberg, 1999; Meunier et al., 2008). Generally, the basal plate is about as thick as the superficial and middle layers combined. However, in *Macropoma lewesiensis* (Williamson, 1849), the basal plate was described as being thinner than these layers, a condition that appears to be variably developed within taxa. On the contrary, in *Latimeria*, the unmineralized basal plate is greatly developed whereas the middle and superficial layers are reduced (Castanet et al., 1975; Meunier et al., 2008) (Fig. 10). In *Miguashaia bureau*, the thickness of the isopedine varies depending on the location of the thin section across the scales: the basal plate appears relatively thicker in the anterior portion (overlapped area) than in the posterior portion (exposed area) of the scales (Figs 3C, 4, 5A, 7A).

The isopedine layer consists of a cellular lamellar bone that can present enclosed osteocytes, as in *Miguashaia bureau* (Figs 4C,D, 7B, 8A), *Spermotodus pustulosus* (Meinke, 1982) and *Undina penicillata* (Gross, 1935, 1966). In *S. pustulosus*, the

isopedine from the dermal bones shows traces of secondary ossification (Meinke, 1982), as in the lateral line scales of *Latimeria chalumnae* (Meunier 1980, figs. 14, 15). Vertical vascular canals can also occur in the scales of *Macropoma lewesiensis* (Williamson, 1849) and in *Miguashaia bureaui* (Fig. 7B). The isopedine of *Miguashaia* has a twisted, plywood-like structure, similar to that of *Latimeria* and possibly other extinct coelacanths with well-developed lamellated basal plates (Fig. 8). However, it differs from the isopedine of *Latimeria* in being fully mineralized and in the lamellar bone being simply twisted, as opposed to the double-twisted arrangement in *Latimeria* (Giraud et al., 1978a,b; Francillon-Vieillot et al., 1990). A double-twisted plywood is also known to occur convergently in other extinct sarcopterygians with rounded scales, like in the tetrapodomorph *Eusthenopteron foordi* (Zylberberg et al., 2010) and the dipnomorph *Holoptychius* sp. (Mondéjar-Fernández & Meunier, 2020), as well as in the living dipnoans *Neoceratodus forsteri*, *Lepidosiren paradoxa* and *Protopterus* (*P. annectens* and *P. aethiopicus*) (Meunier & François, 1980). The twisted condition is thus commonly found in round-scaled sarcopterygians and appears to be derived from a primitive orthogonal arrangement of the isopedine present in stem osteichthyans (e.g. *Psarolepis*) (Qu et al., 2013) and many sarcopterygians with rhombic scales (e.g. *Porolepis*, *Osteolepis*) (Mondéjar-Fernández & Meunier, 2020). However, given the current availability of

paleohistological data on fossil coelacanth material, it is unclear whether a double-twisted plywood is an autapomorphy of *Latimeria* or if it was inherited from closely-related taxa. Finally, it is now well established that the irregularly mineralized basal plate of the scales of *Latimeria* derives from the fully mineralized basal plate of closely-related extinct coelacanths and is topologically and structurally homologous to the isopedine layer from other early sarcopterygians (e.g. early actinistians like *Miguashaia*, dipnomorphs like *Porolepis*, *Holoptychius* and *Dipterus*, and tetrapodomorphs like *Megalichthys* and *Eusthenopteron*) (Mondéjar-Fernández & Meunier, 2020).

Scale development and ontogeny

Information on the development and patterning of the squamation in coelacanths is available through the study of the extant *Latimeria*. Smith (1979) showed that in the scales of *Latimeria* the odontodes develop separately from the basal plate (isopedine) and later fuse to it. In embryos of *Latimeria*, the odontodes are firmly associated with the vascular bone of the spongiosa and these develop synchronously after differentiation of the layers of the basal plate. During growth and development of the scales, supplementary odontodes and more spongy bone are added centrifugally and some occur superimposed onto the earlier generation (Fig. 10A; Roux, 1942; Smith et al., 1972; Ørvig, 1977; Smith, 1979).

Developmental data on *Latimeria* thus matches developmental (Sire & Huysseune, 2003) and evolutionary (Sire et al., 2009) scenarios of the squamation in osteichthyans as the development and evolution of the squamation appear to occur on two levels, matching the two main portions of the scales: 1) the odontogenic component (neural crest-derived) comprising the combination of the superficial odontode-bearing layer and associated vascular bone from the middle layer, and 2) the osteogenic component (mesodermally-derived) represented by the basal plate. The superficial layer and basal plate of the scales of osteichthyans show independent patterning and evolutionary trends, consistent with separate developmental origins (Gillis et al., 2009), a pattern that can be identified from the first manifestations of the dermoskeleton in early vertebrates (Donoghue et al., 2006). Microanatomical and histological changes of tissues (e.g. loss of cosmine, modification of odontode shape and pattern of distribution) can occur in the superficial layer of the scales without affecting the basal plate, and vice-versa (e.g. loss of mineralization of the isopedine), revealing the independence of the cell lineages responsible for the odontogenic and osteogenic components of the dermal skeleton (Schaeffer, 1977).

Unfortunately, little information is available on the development of the squamation in extinct actinistians. Ontogenetic series are fairly rare and embryonic or early

juvenile stages are solely known *Miguashaia bureaui* (Schultze, 1973; Cloutier, 1996; Cloutier et al., 2009), *Serenichthys kowiensis* from the Devonian of South Africa (Gess and Coates, 2015), *Rhabdoderma exiguum* from the Carboniferous of the USA (Schultze, 1972, 1980; Cloutier, 2010), *Caridosuctor populosum*, *Hadronector donbairdi* and *Polyosteorhynchus simplex* from the Carboniferous of the USA (Lund and Lund, 1985), *Luopingcoelacanthus eurylacrimalis* from the Triassic of China (Wen et al., 2013), *Undina penicillata* from the Jurassic of Germany (Watson, 1927; Clack, 1996), *Axelrodichthys* sp. from the Cretaceous of Brazil (Brito and Martill, 1999), and a juvenile coelacanth from the Carboniferous of Germany (Witzmann et al., 2010). The most diagnostic character to recognize juvenile coelacanths is the distinctly longer supplementary lobe of the caudal fin relative to that of adults. *Miguashaia* (Cloutier, 1996) and *Gavinia* (Long, 1999) are considered morphologically primitive coelacanths by the possession of an heterocercal caudal fin lacking a supplementary lobe as opposed to more derived and anatomically modern coelacanths with a characteristic trilobed diphyccercal caudal fin (Zhu et al., 2012). However, in *Miguashaia*, the notochordal lobe is proportionately much longer in juveniles (Cloutier et al., 2009).

Developmental studies on extant osteichthyans (e.g. *Danio*) have shown that the squamation develops gradually across the body, usually starting posteriorly, at the level of the caudal

peduncle, and progresses anteriorly following the lateral line (Sire & Akimenko, 2004), a pattern that can be confidently transposed to fossil ontogenies of extinct taxa, with minor differences (Cloutier, 2010). The date of appearance of the first scales is still unknown in *Latimeria*, but embryos found in the oviducts of the females of *L. chalumnae* can be up to 13 months old before birth, and display two lines of arrested growth (Hureau & Ozouf, 1977). However, the squamation is rarely preserved in juvenile coelacanth fossil specimens as scales are among the latest elements to ossify during the ontogeny, and this usually prevents fossilization. One noteworthy exception is *Rhabdoderma exiguum* for which small yolk-sac embryonic specimens (e.g. YPM 2024, 24.2 mm in standard length; FMNH PF 5521) display extremely thin scales present throughout the complete body and are ornamented with small longitudinal ridges (R.C., pers. observ.). The case of *Miguashaia bureaui* is also remarkable since juvenile as well as adult specimens are known in which the squamation is fully developed and preserved (Schultze, 1973; Cloutier, 1996).

The variation in the ornamentation of the gular plates and scales has been listed among the main ontogenetic changes occurring in *Miguashaia* (Cloutier, 1996). In the case of the scales, the extent of the pitted enamel surface increases during growth, from juveniles with incompletely covered exposed areas in the posterior portion of the trunk to adults displaying a fully sheeted ornamented area (Fig. 2E). It has

also been shown that small individuals mainly display fine longitudinal ridges across the body (ULQ 120, MHNM 06-1633), while coarse tubercles and larger ridges are mainly found in larger individuals (MHNM 06-641). In the small juveniles, the internal surface of the scales lacks the small circular bumps which become visible only in adult specimens (Fig. 2D2).

Ornamentation variation across the ontogeny is also known in *Rhabdoderma exiguum* in which the ornamentation of posterior scales is more pronounced than in the anterior part of the body (e.g. FMNH PF 5760) and the number of longitudinal ridges increases from embryonic to juvenile specimens.

Remodeling of bone, which produces structures such as the secondary vascular canals in *Miguashaia bureaui* (Fig. 4E, 6), is also evidence of advanced ontogenetic growth (Dias & Richter, 2002) and may be related to sexual maturity in these specimens.

The development of larger tubercles and ridges has been associated with the function of tubercles as a reinforcement of the adhesion properties between the scales and the overlying epithelium (Burdak, 1979). The shape and localization of these tubercles across the body are also related to the control of the water flow during swimming throughout ontogeny, as evidenced in extant osteichthyans (e.g. *Mugil*) (Burdak, 1979). The ornamentation in *Latimeria* does not vary significantly during ontogeny, with the only exception of the number of tubercles, which are less numerous

in juvenile specimens due to the small size of the scales, and more abundant in adult specimens with larger scales.

Growth rings (*annuli*) are frequently encountered in the scales of extant and extinct osteichthyans and they occur in the overlapped area of the scales of many early coelacanths (Fig. 11) and of *Latimeria*. These *annuli* have been counted in *L. chalumnae* (Hureau & Ousouf, 1977; Millot & Anthony, 1978), where juvenile specimens have fewer *annuli* than adult ones, and assuming that each ring represents a seasonal or annual line of arrested growth, a putative age can be assigned to specimens based on isolated scales. In *L. chalumnae*, two episodes of arrested growth (January-February and August-September) occur each year during the dry season (Hureau & Ozouf, 1977), allowing to assign a relatively accurate age to specimens caught in the wild. In *Miguashaia bureaui* equally spaced *annuli* are present in large adult specimens (e.g. MHNM 06-641, 06-1232, 06-1236, 06-1239, 06-1809) and are more numerous than in smaller individuals. P; peripheral growth rings are more closely spaced towards the periphery of large scales, congruent with the acquisition of sexual maturity and a slower growth rate in adults. Approximative ages could also be given to fossil specimens but these estimations can be less reliable given the tentative reconstruction of the paleoenvironnements in which these forms lived and the uncertainty regarding the metabolism, food supply, competitive

stress, salinity and duration of the seasons, among other possible factors.

Evolutionary trends of the squamation in coelacanth

Among other synapomorphies, coelacanth are characterized by the possession of rounded scales (Janvier, 1996; Forey, 1998). However, the squamation of actinistians is rarely composed of perfectly circular scales. The scales can either be roughly circular (e.g. in early forms like *Miguashaia* and *Diplocercides*) or subcircular (oblong to oval), with a slightly quadrangular anterior margin (e.g. *Guizhoucoelacanthus*, *Latimeria*). Other taxa possess ovate scales, with a pronounced posterior edge that may display a rounded (e.g. *Sassenia*, *Whiteia*) or pointed margin (e.g. *Lualabea*, *Diplurus*, *Rhaboderma*). Given the morphological plasticity of the squamation, the body size variation, and the relative environmental cosmopolitanism of coelacanth across their evolutionary history, no distinctive trend can be depicted for the evolution of the scale outline; however, the earliest taxa (e.g. *Miguashaia*, *Diplocercides*) display the most circular contour whereas in more recent forms the scales become more antero-posteriorly elongated and the posterior margin displays a more variable profile, with different degrees of acuteness (Fig. 11).

The size, extension, and combination pattern of tubercles and ridges that occur on the exposed area is also extremely

variable (Forey, 1998). In many genera, the size and distribution of the ridges and tubercles are homogeneous (e.g. *Rhabdoderma*, *Caridosuctor*, *Holophagus* and *Latimeria*) but some taxa display a differentiated ornament in which large ridges usually located in the central portion of the exposed area are associated with smaller ridges that are distributed laterally (e.g. *Axelrodichthys*, *Chinlea*, *Diplurus*, *Heptanema*, *Lualabea*, *Mawsonia*) (Forey, 1998, Yabumoto, 2008; Renesto & Stockar, 2018; Fragoso et al., 2019). This condition appears to be especially developed in the Mawsoniidae (Fig. 11) and may constitute a shared derived feature of these taxa but further phylogenetic inquiry is needed to confirm it.

~~Ornamental~~ variations of the ornamentation have been used to discriminate taxa and thus have a taxonomic value at generic or species levels (e.g. various species of *Rhabdoderma* and *Whiteia*) (Table 1). In completely preserved specimens, ornamentation variation has been observed to occur over various regions of the body (e.g. *Miguashaia*, *Macropoma*, *Luopingcoelacanthus*) (Cloutier, 1996; Forey, 1998; Wen et al., 2013) (Fig. 2) and thus taxonomic identification must be cautious when describing new taxa based solely on isolated scale remains. A general evolutionary pattern nevertheless emerges, in which the earliest coelacanth (e.g. *Miguashaia*, *Diplocercides*) tend to show numerous tubercles or ridges, whereas younger taxa (e.g. *Rhabdoderma*, *Diplurus*, *Latimeria*) display fewer but larger tubercles and ridges (Figs 2C, 11).

By contrast, the overlapped area of the scales is not ornamented, excepted in some Devonian taxa like *Miguashaia*, *Gavinia*, and *Diplocercides*, which display a reduced field of small leaf- or spoon-shaped tubercles spanning the margin between the overlapped and exposed areas (Figs 2C-E, 11). These small odontodes are also characteristic of other Devonian sarcopterygians (e.g. onychodonts, porolepiforms, early dipnoans), where they may occur associated with a well-developed cosmine sheet (e.g. the porolepiforms *Porolepis*, *Heimenia*, and the early dipnoan *Uranolophus*) (Ørvig, 1957, 1969; Denison, 1968; Mondéjar-Fernández & Clément, 2012) or not (e.g. the porolepiforms *Holoptychius*, *Glyptolepis*, and *Laccognathus*, and the onychodonts *Onychodus* and *Selenodus*) (Ørvig, 1957; Cloutier & Schultze, 1996; Andrews et al., 2006; Mondéjar-Fernández, 2020). Such small odontodes thus represent a primitive shared feature of sarcopterygians. Past the Carboniferous, coelacanth scales lost this type of tubercles, yielding a relatively smooth overlapped area and fewer and larger tubercles and ridges in the exposed area (e.g. *Caridosuctor*, *Rhabdoderma*, *Axelrodichthys*) (Fig. 11).

The overlapped area generally occupies between a half and two thirds of the external surface of the scales of coelacanths. However, the isolated scale material is scarce in many post-Devonian taxa and thus neither the smoothness nor the precise contour of the overlapped area can be properly described or illustrated in these taxa (see Forey, 1998). In

the scales from the posterior part of the trunk of *Miguashaia bureaui*, the ornamented exposed area is almost as developed as the overlapped area with a roughly 50/50 ratio (Fig. 2B,E), most probably representing the primitive condition in actinistians. Other Devonian taxa such as *Diplocercides* also display an important exposed area relative to the overlapped area (Fig. 11). However, the anterior portion of the overlapped area is slightly more developed and tend to widen relative to a narrower posterior portion of the exposed area, a pattern that is accentuated in post-Devonian forms (e.g. *Diplurus*, *Axelrodichthys*, *Guizoucoelacanthus*, *Latimeria*).

The internal surface of the scales is poorly-known in extinct coelacanth as few specimens preserve the scales in internal view. To our knowledge, the internal surface has solely been described among Palaeozoic coelacanth in *Miguashaia bureaui*, where small numerous bony bumps occur in the posterior region (Fig. 2D2). In Mesozoic forms, the internal surface of the scales is preserved in the Triassic *Luopingcoelacanthus eurylacrimalis* (Wen et al., 2013), the Jurassic *Indocoelacanthus robustus* (Jain, 1974), and the Cretaceous *Megalocoelacanthus dobiei* (Dutel et al., 2012) where it is smooth but concentric growth rings are well developed. The scales of the extant *Latimeria* have a smooth, slightly concave but otherwise featureless, internal surface. *Foreya maxkuhni* from the Triassic (Cavin et al., 2017) is an interesting case as numerous scales from the trunk and base of

the fins are known. The internal surface is generally smooth but in some portions of the body a drop-shaped boss is clearly visible (Cavin et al., 2017, fig. 2.4,5,8). This feature is reminiscent of the distinctive drop-shaped boss of certain Devonian sarcopterygians like rhizodonts and tristichopterids (Jarvik, 1980). The phylogenetic history and significance of this character is difficult to retrace. Its occurrence in *Foreyia* may represent the result of a simplification of the pattern seen in *Miguashaia* with numerous bumps or an independently acquired convergent feature with Devonian tetrapodomorphs.

The early evolution of the coelacanth scales

Two enigmas remain in the early evolution of ~~coelacanth's~~ the squamation in coelacanths: 1) did coelacanths lose cosmine (as in many other sarcopterygian groups during the Devonian), or was cosmine never present in their ancestors? and 2) was the primitive scale morphotype of coelacanths rhombic or rounded? Indeed, *Miguashaia* and *Gavinia*, the currently considered basal-most undisputed coelacanths (Zhu et al., 2012; Gess & Coates, 2015) display rounded scales without cosmine and ornamented with the classical odontogenic combination of tubercles and ridges. The position of the putative actinistian *Styloichthys* and the likely sister-group relationship between coelacanths and onychodonts among early sarcopterygians are

key to answer these questions, however both issues are still unsettled.

Styloichthys changeae from the Lochkovian (Lower Devonian) of China was initially described as the sister-group of rhipidistians (defined as the largest clade that includes all taxa more closely related to tetrapods and diponans than to actinistians) (Zhu & Yu, 2002) and this position has been subsequently confirmed in other analyses (e.g. Zhu et al., 2006, 2009; Lu et al., 2016a,b). However, after Friedman's (2007) new interpretation, its status as a putative early coelacanth has started to gain acceptance (e.g. Gess & Coates, 2015; Lu et al., 2017). *Styloichthys* displays shared derived characters of the lower jaw, braincase and dermal skull with coelacanths (Friedman, 2007), however, as opposed to other actinistians, *Styloichthys* presents rhombic cosmoid scales (Lu & Zhu, 2008; Cui et al., 2019), similar in shape to those of early dipnomorphs (e.g. *Youngolepis*) (Chang & Smith, 1992) and tetrapodomorphs (e.g. *Kenichthys*) (Chang & Zhu, 1993). Cross sections performed in the dermal shoulder girdle (cleithrum), but not in the scales, show that *Styloichthys* possesses a peculiar type of cosmine in which large odontodes are associated with a pore-canal system that opens to the surface through wider pores than in rhipidistians (i.e., dipnomorphs and tetrapodomorphs) but similar to other early osteichthyans (e.g. *Psarolepis* and *Meemannia*) (Zhu et al., 2006; Qu et al., 2013; Mondéjar-Fernández, 2018). In *Styloichthys*, the pore-

canal system is embedded in several layers of enamel and dentine, organized as superimposed odontodes (Zhu et al., 2006, 2010). This condition differs from the classical "true" cosmine (eucosmine) of rhipidistians where a single layer of enamel overlays the dentine (Sire et al. 2009; Mondéjar-Fernández 2018) and highlights that *Styloichthys* exhibits a primitive type of cosmine, similar to that of stem osteichthyans like *Psarolepis* and *Achoania*, and early actinopterygians like *Meemannia* (Zhu et al., 2006, 2010; Qu et al., 2013, 2016; Lu et al., 2016b).

The case of onychodonts is also of interest when dealing with the primitive condition of the squamation in actinistians. A close relationship between onychodonts and coelacanth has been proposed in numerous recent studies of sarcopterygian interrelationships (e.g. Zhu et al., 1999, 2001, 2006, 2009; Botella et al., 2007; Lu & Zhu 2010; Lu et al., 2016b, 2017; Clement et al., 2018) and it is currently suspected that onychodonts might represent a paraphyletic group of stem actinistians (Lu et al., 2016a; Mondéjar-Fernández, 2020). However, this result complicates the reconstruction of the primitive scale morpho- and histotype in coelacanth. The scales of all known onychodonts are rounded, except in *Qingmenodus yui* from the Pragian (Lower Devonian) of China, for which the scale shape is still unknown (Lu & Zhu, 2010; Lu et al., 2016a). The exposed area of the scales is classically ornamented with characteristic pointed tubercles,

confirmed to be odontodes made of dentine capped with a thin enamel layer in *Selenodus aquesbiae* from the Eifelian (Middle Devonian) of Morocco (Mondéjar-Fernández, 2020). In the earliest undisputed onychodont *Bukkanodus jesseni* from the Pragian of Australia (Johanson et al., 2007), the exposed area is ornamented with wide dentine ridges separated by grooves containing pores. The odontodes of onychodonts are histologically identical to those of coelacanth-like *Latimeria* (Castanet et al., 1975; Meunier et al., 2008), and now also confirmed in *Miguashaia* (Figs 3,4), which probably represents a primitive shared condition of onychodonts and actinistians. Absence of cosmine is also a diagnostic feature ~~for~~of onychodonts (Andrews et al., 2006). However, putative onychodont remains from the Lower and Middle Devonian of China are covered with cosmine (Zhu & Yu, 2004; Zhu & Zhao, 2005; Lu & Zhu, 2010). The dermal skull bones of *Qingmenodus* display a shiny coating, probably made of enamel, and closely spaced tiny pores, but no histological information is available to confirm or refute the occurrence of cosmine (Lu & Zhu, 2010; Lu et al., 2016a). It would thus not be surprising that new material from early onychodonts present cosmine, which would suggest that the cosmine was lost in younger onychodonts and actinistians.

The current phylogenetic picture of coelacanth interrelationships within sarcopterygians evidences two problems that complicate the unravelling of the origin of the

coelacanth squamation (Fig. 12A). If *Styloichthys* is considered as the oldest coelacanth and the sister-group of all other actinistians (Friedman, 2007; Gess & Coates, 2015), then 1) a rhombic and cosmoid morpho- and histotype may be primitive for coelacanths, which implies that 2) the acquisition of a rounded shape and the loss of cosmine probably occurred during the Lower-Middle Devonian, as displayed in *Miguashaia*, *Gavinia* and *Diplocercides* (Fig. 11). However, this scenario is at odds with the supposed sister-group relationship of onychodonts and coelacanths, especially if onychodonts are no longer considered a monophyletic group but a subset of stem actinistians (Lu & Zhu, 2010; Lu et al., 2016a,b, 2017; Mondéjar-Fernández, 2020). In this latter case, the evolution of a cosmoid rhombic morphotype of *Styloichthys* from the rounded and cosmine-lacking scales of onychodontids implies a reversal (Fig. 12B).

A series of plausible scenarios could reconcile the phylogenetic position of *Styloichthys* with the close relationship of onychodonts and coelacanths and parsimoniously account for the evolutionary origin of the actinistian squamation. The position of *Styloichthys* has been reconstructed as shifting between the sister-group of the onychodont-actinistian clade (Lu et al., 2016b) (Fig. 12C) and the sister-group of a more inclusive taxon that includes the onychodont-actinistian clade and rhipidistians (Lu et al., 2017) (Fig. 12D). If either of these hypotheses are further

supported by new fossil and phylogenetic data, then it will corroborate that the combination of a rounded scale morphotype and the ridged/tuberculated ornamentation of onychodonts and coelacanths is a shared feature. Rounded scales devoid of cosmine might have evolved from primitive cosmoid scales similar to those of early sarcopterygians like *Styloichthys* (Fig. 12C) or from early cosmine-covered onychodonts (Fig. 12D) before the establishment of the rounded morphotype in the common ancestor of more recent onychodonts (like *Bukkanodus*, *Selenodus*, *Onychodus* or *Strunius*) and actinistians.

The last possible scenario is to consider onychodonts as stem sarcopterygians, not closely related to actinistians, but rather as the sister-group of a clade comprising coelacanths (including *Styloichthys*) and rhipidistians (e.g. Cloutier & Ahlberg, 1996; Friedman, 2007) (Fig. 12E). In this case, the acquisition of a rounded and tuberculated squamation is convergent in onychodonts and actinistians, and both originated from a rhombic ancestral morphotype covered in cosmine, similar that of stem osteichthyans like *Psarolepis* (Qu et al., 2013). In the case of onychodonts, primitive cosmoid forms may be represented by the putative cosmine-covered onychodonts from China (Zhu & Yu, 2004; Zhu & Zhao, 2005; Lu & Zhu, 2010), whereas for coelacanths, the primitive condition may be exemplified by the rhombic scales of *Styloichthys* (Zhu & Yu, 2002; Zhu et al., 2006, 2010; Friedman, 2007).

CONCLUSION

The scales of the early coelacanth *Miguashaia bureaui* from the Frasnian (Upper Devonian) of Miguasha, Québec (Canada), have been histologically investigated ~~for the first time~~, revealing a combination of primitive features for actinistians that allow us to reconstruct the primitive condition of the squamation of coelacanths. The rounded scales devoid of cosmine of *Miguashaia* can be described as elasmoid scales, in which the superficial layer is ornamented by pointed tubercles and narrow ridges made of dentine capped with enamel (odontodes). At least two generations of superimposed odontodes occur, reminiscent of the primitive condition of stem osteichthyans like *Andreolepis* or *Lophosteus*. The middle vascular layer (spongiosa) is well developed and tightly linked to the odontodes. The basal plate (isopedine) consists of a fully mineralized layer of lamellar bone with a twisted plywood-like arrangement of the collagen plies, which display a repetitive rotation pattern every five layers.

The presumably primitive condition of the scales of *Miguashaia* allows testing evolutionary scenarios for the squamation in coelacanths by comparing *Miguashaia* with other extinct (e.g. *Diplocercides*, *Diplurus*) and extant (*Latimeria*) actinistians. Despite an important lack of paleohistological comparative material from the scales of extinct coelacanths,

several evolutionary trends can be inferred from Palaeozoic taxa to Recent: 1) a reduction in number but increase in size of the tubercles and ridges ornamenting the exposed area, 2) an antero-posterior elongation of the scales, 3) the development of larger pulp cavities in the dentine forming the odontodes, and 4) an increase in the relative thickness of the basal plate, coupled with the reduction of the mineralization rate of the isopedine in *Latimeria*.

Overall, the combination of traits described in *Miguashaia* do not represent a drastic departure from the histological structure of the scales of other coelacanths like *Macropoma* (Williamson, 1849), *Coelacanthus* (Goodrich, 1907), or *Undina* (Gross, 1935, 1966). Coelacanths are considered to have experienced an early burst of diversification during the Palaeozoic, followed by remarkable morphological conservatism with limited variations until Recent (e.g. Cloutier, 1991a). The scales of *Miguashaia* already possess the more characteristic features of the actinistian squamation, consistent with this general evolutionary scenario. However, the current uncertainties regarding the position of the putative coelacanth *Styloichthys* and the role of onychodonts as the sister-group of actinistians complicates the reconstruction of the onset of the coelacanth scales prior to the emergence of early ~~coelacanths~~-forms like *Miguashaia* or *Gavinia*. Additional paleohistological data are thus needed from other extinct actinistians and early sarcopterygians.

These histological features could then be incorporated into future phylogenetic analyses, which would contribute to the unravelling of actinistian interrelationships and continue shedding more light on the evolutionary history of the squamation of coelacanths.

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AUTHORS CONTRIBUTION

JMF, FJM and GC designed the study. ML provided the fossil material. JMF and FJM acquired the histological cross sections and analysed the data. RC completed the morphological observations. JMF prepared the figures and table, and wrote the text with contributions from FJM and RC. All authors revised the manuscript and approved its final version.

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TABLES

Table 1. List of extinct coelacanths for which the squamation has been studied and figured.

Genus	Species	Scale Morphology			Scale Histology				Reference	
		Shape	External surface		Internal surface	Enamel/enameloid	Dentine	Vascular bone		Lamellar bone
			Exposed area	Overlapped area						
<i>Allenypterus</i>	<i>A. montanus</i>	spindle-shaped (ventromedian scales)	unnamed?	–	–	–	–	–	–	Lund & Lund, 1985
<i>Axelia</i>	<i>A. robusta</i>	ovate	stout ridges and tubercles	–	–	–	–	–	–	Stensiö, 1921
<i>Axelrodichthys</i>	<i>A. araripensis</i>	ovate	long central ridge and narrow lateral ridges	–	–	–	–	–	–	Maisey, 1986; Fragoso et al., 2019

<i>Caridosuctor</i>	<i>C. populosum</i>	ovate	narrow ridges	-	-	-	-	-	-	Lund & Lund, 1985; Forey, 1998
<i>Chagriina</i>	<i>C. enodis</i>	oval	unornamented	-	-	absent?	absent?	present	-	Schaeffer, 1962
<i>Changxingia</i>	<i>C. aspratilis</i>	oval	elongate ridges	-	-	-	-	-	-	Wang & Liu, 1981
<i>Chaohuichthys</i>	<i>C. majiashanensis</i>	ovate	narrow ridges	-	-	present?	-	-	-	Tong et al., 2006
<i>Chinleia</i>	<i>C. sorensoni</i>	oval	narrow ridges	concentric growth rings	-	-	-	-	-	Schaeffer, 1967
<i>Coccoderma</i>	<i>C. substriolatum</i>	oval	small ridges	-	-	-	-	-	-	Forey, 1998
	<i>C. suevicum</i>		narrow ridges	-	-	-	-	-	-	Forey, 1998
<i>Coelacanthus</i>	<i>C. granulosus</i>	oval	tubercles	-	-	absent?	absent? large pul	osteonal remode	fully mineralised	Goodrich, 1907; Forey

							p cav iti es?	lli ng		ey, 199 8
	<i>C. lunze nsis</i>		elon gate ridg es	-	-	-	-	-	-	For ey, 199 8
<i>Diploc ercide s</i>	<i>D. heili genst ocken sis</i>		elon gate ridg es	thi n rad ial rid ges	-	-	-	-	-	Jes sen , 196 6, 197 3
	<i>D. jaeck eli</i>	subc ircu lar	shor t ridg es and spoo n- shap ed tub ercl es	thi n rad ial rid ges	-	prese nt?	pre sen t?	-	-	Ste nsi ö, 192 2, Gro ss, 193 5, 196 6
	<i>D. kayse ri</i>		elon gate ridg es	thi n rad ial rid ges	-	-	-	-	-	Ste nsi ö, 193 7, For ey, 199 8
<i>Diplur us</i>	<i>D. newar ki</i>	ovat e	elon gate stou t ridg es	-	-	prese nt?	pre sen t?	pre sen t?	ful ly min era lis ed?	Sch aef fer , 195 2; For ey, 199 8
	<i>D. longi cauda tus</i>		elon gate narr ow	thi n rad ial	-	prese nt?	-	-	-	Sch aef fer ,

			ridges	ridges						1948
<i>Foreyia</i>	<i>F. maxkuhni</i>	oval	stout ridges or tubercles and fine striations posteriorly	concentric growth rings	smooth, drop-shaped bosses	-	-	-	-	Cavin et al., 2017
<i>Gavini</i>	<i>G. syntrips</i>	subcircular	wavy ridges surrounded by finer ridges and spoon-shaped tubercles	-	-	-	-	-	-	Long, 1999
<i>Garnbergia</i>	<i>G. ommat</i>	oval	narrow ridges	-	-	-	-	-	-	Martin & Wenz, 1984
<i>Graphiurichthys</i>	<i>G. callopterus</i>	oval	tubercles	-	-	-	-	-	-	Kner, 1866
<i>Guizoucoelacanthus</i>	<i>G. guanl</i>	oval - quad	stout ridge	concentric	-	-	-	-	-	Liu et al.

	<i>ingen sis</i>	rang ular	es and elongate tubercles	c growth rings						, 2006 ; Gen g et al. , 2009
<i>Hadron ector</i>	<i>H. donba irdi</i>	oval	coar se ridg es	-	-	-	-	-	-	Lun d & Lun d, 1984
<i>Hainbe rgia</i>	<i>H. granu lata</i>	oval	narr ow ridg es	-	-	-	-	-	-	Sch wei zer , 1966
<i>Heptan ema</i>	<i>H. parad oxum</i>	oval	prom inen t medi an ridg e and smal ler late ral ridg es	-	-	-	-	-	-	Bel lot i, 1857; For ey, 1998
	<i>H. sp</i>			-	-	-	-	-	-	Ren est o & Sto cka r, 2018
<i>Holoph agus</i>	<i>H. gulo</i>	oval	elon gate ridg es and tubercles	-	-	-	-	-	-	For ey, 1998
<i>Holopt erygiu s</i>	<i>H. nudus</i>	spin dle-shap ed	unor name nted ?	-	-	-	-	-	-	Jes sen ,

		(ventromedian scales)								1973; Friedman & Coates, 2006
<i>Indocoelacanthus</i>	<i>I. robustus</i>	oval	small ridges and striations	concentric growth rings	concentric growth rings	-	-	-	-	Jain, 1974
<i>Laugia</i>	<i>L. groenlandica</i>	ovate	flattened ridges	-	-	-	-	-	-	Stensiö, 1932
<i>Libys</i>	<i>L. polyp terus</i>	oval	short ridges	-	-	-	-	-	-	Münster, 1842
<i>Lualabaea</i>	<i>L. henryi</i>	ovate	coarse ridges	-	-	-	-	-	-	Saint-Seine, 1955; Forey, 1998
<i>Luopingcoelacanthus</i>	<i>L. eurylacrimalis</i>	ovate	stout ridges and elongate tubercles	thin radial ridges?	smooth	-	-	-	-	Wen et al., 2013
<i>Macropoma</i>	<i>M. lewes</i>	oval	large	concentric	-	enamel	orthod	osteon	fully	Wille

	<i>iensis</i>		tubercles	triangular growth rings			entire, large pulp cavities	al remodeling	mineralized	mason, 1849; Forey, 1998
	<i>M. precursor</i>			-	-	-	-	-	-	Forey, 1998
	<i>M. speciosum</i>			-	-	-	-	-	-	Forey, 1998
<i>Macropomoides</i>	<i>M. orientalis</i>	oval	large tubercles	-	-	-	-	-	-	Forey, 1998
<i>Mawsonia</i>	<i>M. brasiliensis</i>	oval	unornamented	concentric growth rings	-	absent?	absent?	present	-	Yabumoto, 2002
<i>Megalocoelacanthus</i>	<i>M. dobie</i>	oval	-	concentric growth rings	-	-	-	-	-	Dutel et al., 2012
<i>Miguashaia</i>	<i>M. bureau</i>	subcircular	numerous tubercles, small ridges and fine striations	thin radial ridges, concentric growth rings	concentric growth rings and bumps	enamel	orthodontine, small pulp cavities,	osteonal remodeling, embedded	fully mineralized	this study

			ations posteriorly	with rings, and spoon- shaped tubercles	teriorly		superimposed odontodes	ntodes		
<i>Mylacanthus</i>	<i>M. lobatus</i>	oval	small coarse ridges	-	-	-	-	-	-	Stensiö, 1921
<i>Parnai baia</i>	<i>P. maranhensis</i>	ovate	prominent median ridges and smaller lateral ridges	-	-	-	-	-	-	Yabumoto, 2008
<i>Pivetea</i>	<i>P. madagascariensis</i>	oval	elongate ridges	-	-	-	-	-	-	Clément, 1999
<i>Rebellatrix</i>	<i>R. divaricata</i>	oval	narrow ridges	-	-	-	-	-	-	Wendru f & Wilson, 2012
<i>Rhabdo derma</i>	<i>R. ardrossense</i>	oval	posteriorly convergent ridges	-	-	-	-	-	-	For ey, 1998
	<i>R. elegans</i>			-	-	-	-	-	-	For ey,

			es							1998
	<i>R. exiguum</i>			-	-	-	-	-	-	For ey, 1998
	<i>R. huxleyi</i>			-	-	-	-	-	-	For ey, 1998
	<i>R. madagascariensis</i>			-	-	-	-	-	-	For ey, 1998
	<i>R. tinglensis</i>			-	-	-	-	-	-	For ey, 1998
<i>Sassenia</i>	<i>S. groenlandica</i>	oval	tubercles and small ridges	-	-	-	-	-	-	For ey, 1998
<i>Sassenia</i>	<i>S. tuberculata</i>			-	-	-	-	-	-	Stensi ö, 1932
<i>Scleranthus</i>	<i>S. asper</i>	ovate	coarse, sharp ridges	-	-	-	-	-	-	Stensi ö, 1921
<i>Serenichthys</i>	<i>S. kowieensis</i>	oval	elongate ridges	-	-	-	-	-	-	Gess & Coates, 2015
<i>Sinocoelacanthus</i>	<i>S. fengshanensis</i>	oval	-	concentric growth rings	-	-	-	-	-	Liu, 1964
<i>Spermotodus</i>	<i>S. pustulosus</i>	oval	tubercles	-	-	present	present	present	-	Westoll,

						(dermal bones)	(dermal bones)	(dermal bones)		1939: Meinke, 1982
<i>Swenzia</i>	<i>S. latimerae</i>	oval	elongate tubercles	-	-	-	-	-	-	Clément, 2005
<i>Ticine pomis</i>	<i>T. cf. T. peyeri</i>	oval	elongate ridges	-	-	-	-	-	-	Cavin et al., 2013
<i>Trachymetopon</i>	<i>T. liassicum</i> ,	oval	elongate ridges	-	-	-	-	-	-	Dutel et al., 2015
<i>Undina</i>	<i>U. cirinensis</i>	oval	elongate tubercles and ridges	-	-	-	-	-	-	Saint-Seine, 1949
	<i>U. penicillata</i>			-	-	absent	orthodontine, large pulp cavities	osteonal remodeling? embedded odontodes	-	Münster, 1834; Gross, 1935, 1966
	<i>U. purbeckensis</i>			-	-	-	-	-	-	Woodward, 1916

<i>Whiteia</i>	<i>W. nielsenii</i>	oval	short ridges	-	-	-	-	-	-	For ey, 1998
	<i>W. oishi</i>		elongate ridges	-	-	-	-	-	-	Yab umoto & Bri to, 2016
	<i>W. tuberculata</i>		tubercles	-	-	-	-	-	-	For ey, 1998
	<i>W. uyeneruyai</i>		elongate ridges	-	-	-	-	-	-	Yab umoto et al., 2019
	<i>W. woodwardi</i>		elongate tubercles	-	-	-	-	-	-	For ey, 1998
<i>Wimania</i>	<i>W. sinuosa</i>	ovate	short ridges	concentric growth rings	-	-	-	-	-	Ste nsi ö, 1921
<i>Youngiichthys</i>	<i>Y. xinhuaisinsis</i>	oval	narrow ridges	-	-	-	-	-	-	Wan g & Liu, 1981

FIGURES

FIGURE 1. Photographs of the fossil specimens of *Miguashaia bureau*i histologically surveyed. **A.** MHNM 06-1238. **B.** MHNM 06-1495. **Abbreviations:** cl, cleithrum; lp, lepidotrichia; n.ar, neural arches; op.s, opercular series; ra, radials, sc, scales. Scale bar equals 10 mm.

FIGURE 2. Squamation of *Miguashaia bureau*i. **A.** Complete specimen (MHNM 06-494). **B.** Reconstruction and body outline of *M. bureau*i with the scales from three locations of the left flank highlighted. **C-E.** Reconstruction of different scales from the left flank: **C.** Scale from the anteroventral portion of the trunk, posterior to the pectoral fin, in external view, **D.** Scale from the posterior portion of the trunk, in external (1) and internal (2) view. **E.** Scale from the posterior-most region of the trunk, close to the caudal fin, in external view. Drawings based on MHNM 06-6, MHNM 06-641 (Cloutier, 1996; fig. 17), MHNM 06-1238, MHNM 06-1239, and MHNM 06-1495. **Abbreviations:** ci.b, circular bumps; co.tu, coarse tubercles; co.ri, coarse ridges; f.ri, fine ridges; pi.e, pitted enamel; sp.tu, spoon-shaped tubercles. Arrow points anteriorly. Scale bars equal 10 cm (**A**) and 10 mm (**C-E**).

FIGURE 3. General microstructural organization of the scales of *Miguashaia bureau*i (vertical cross sections of MHNM 06-

1495). **A.** Superimposed odontodes from the superficial layer. **B.** Vascular bone (spongiosa) from the superficial layer. Arrow heads point to osteocytes embedded in the bone. **C.** Lamellar bone (isopedine) from the basal plate. Black arrows indicate the repetitive pattern of collagenous plies with the same orientation. **Abbreviations:** **d1-2**, dentine layer of two generations of odontodes; **e1-2**, enamel layer of two generations of odontodes; **lb**, lamellar bone; **pc**, pulp cavity; **vb**, vascular bone. Scale bar equals 50 μ m.

FIGURE 4. Ornamentation and microstructural organization of the exposed area of the scales of *Miguashaia bureaui* (vertical cross sections of MHN 06-1495). **A.** General view of a scale showing the distribution of the odontodes in the superficial layer. White arrow heads indicate the irregular front of mineralization of the ventral surface of the basal plate. **B.** Detail of an odontode composed of a thin layer of enamel and dentine with numerous odontoblastic canaliculi overlying the layer of vascular bone (spongiosa). **C.** Inset of two odontodes from **A.** Note the opening of vascular canals from the spongiosa through pores in the surface, as in **B**, and the large odontoblastic lacunae. Arrow heads indicate the occurrence of osteocyte lacunae in the lamellar bone (isopedine) of the basal plate. **D-E.** Different distributions and arrangements of two generations of superimposed odontodes. Note the reduced thickness of the spongiosa and the occurrence of vascular

canals and vascular bone surrounding the older odontodes.

Abbreviations: **bp**, basal plate; **d**, dentine; **d1-2**, dentine layer of two generations of odontodes; **e**, enamel; **e1-2**, enamel layer of two generations of odontodes; **lb**, lamellar bone; **ml**, middle layer; **oc**, odontoblastic canaliculi; **od1-2**, two generations of odontodes; **vb**, vascular bone; **vc**, vascular canal;. Scale bars equal 100 μm .

FIGURE 5. Ornamentation and microstructural organization of the overlapped area of the scales of *Miguashaia bureaui* (vertical and horizontal cross sections of MHNM 06-1238). **A.** Three scales vertically sectioned at different regions of the overlapped area (from anterior to posterior): 1) Anterior most margin; 2) Central region; 3) Posterior region, close to the contact with the exposed area. Note the increase in thickness of the vascular bone layer (spongiosa) and the lamellar bone layer (isopedine) from anterior to posterior and the changes in the bony ornamentation. Scale bar equals 500 μm . **B.** Bony ornamentations of the overlapped area from the anterior most margin (1) and the central region (2). Scale bar equals 100 μm . **C.** Horizontal cross section through the small bony tubercles of the overlapped area from the anterior most margin of a scale (1). Note the concentric patterns of ossification, the radial arrangement of osteocyte lacunae, and the openings of vascular canals between the tubercles. Scale bar equals 50 μm . **Abbreviations:** **lb**, lamellar bone; **vb**, vascular bone.

FIGURE 6. Microstructural and histological organization of the vascular bone of the scales of *Miguashaia bureaui* (horizontal cross sections of MHNM 06-1238). **A.** General view of the vascular bone layer (spongiosa) from the overlapped area of the scales. Note the uniform diameter and distribution of the vascular canals. Scale bar equals 500 μm . **B.** Detailed inset from **A.** Note that each vascular canal is surrounded by a series of concentric layers of woven-fibered bone (light brown) while each row of vascular canals is separated from the other by a denser bone tissue (dark brown). Scale bar equals 100 μm . **C.** Detailed inset from **B.** Arrow heads indicate the occurrence of osteocyte lacunae surrounding the vascular canals. Scale bar equals 50 μm . **D.** General view of the vascular bone and vascular canals from another region of the overlapped area of a scale, probably from a more posterior region than in **A.** Note the more irregular distribution of the vascular canals and the occurrence of ossification waves. Scale bar equals 100 μm . **E.** Detailed inset from **D.** Note the occurrence of large osteocyte lacunae in the woven-fibered bone around the vascular canals. Scale bar equals 50 μm . **F.** Detailed inset from **D.** Arrow heads point to embedded osteocyte lacunae with large osteocytic prolongations. Scale bar equals 50 μm .

FIGURE 7. Microstructural and histological organization of the basal plate of the scales of *Miguashaia bureaui* (vertical cross sections of MHNM 06-1238 and MHNM 06-1495). **A.** General view of the vascular bone layer (spongiosa) of the superficial layer and lamellar bone (isopedine) of the basal plate from the anterior portion of the overlapped area of the scales (MHNM 06-1238). The horizontal plane corresponds to the approximative cross section plane in Fig. 6D-F. Note the clear differences in bone structure between the spongiosa (made of woven-fibered bone) and the isopedine (made of lamellar bone). Scale bar equals 100 μm . **B.** Detailed view of the basal plate from the central portion of the exposed area of the scales (MHNM 06-1495). Black arrows indicate the repetitive pattern of collagenous plies with the same orientation. White arrow heads indicate the irregular front of mineralization of the visceral surface of the basal plate. The black arrow head points to a vertical vascular canal piercing the basal plate and opening through the ventral surface of the scale. Note that the thickness of the basal plate is more important around the vascular canal resulting in a bump of the internal surface of the scale at this point. Scale bar equals 100 μm .

Abbreviations: **bp**, basal plate; **lb**, lamellar bone; **sl**, superficial layer; **vb**, vascular bone; **vc**, vascular canal.

FIGURE 8. Basal plate comparison of the scales of *Miguashaia* and *Latimeria* in horizontal cross section. **A.** *Miguashaia*

bureaui (MHNM 06-1238). Note the occurrence of 5 directions of the collagen fibers. **B.** *Latimeria chalumnae* (MNHN-ZA-AC-2012-29). **C.** *Latimeria menadoensis* (MZB 10003). Note the arciform pattern of the collagen plies evidencing a twisted-plywood arrangement of the layers of the isopedine. Scale bars equal 100 μm .

FIGURE 9. Odontodes from the exposed area of the scales of *Latimeria* (SEM). **A.** *Latimeria menadoensis* (MZB 10003). General view of the odontodes. The posterior margin of the scale is on the upper left of the image. The radial ridges of the superficial layer are clearly seen between the odontodes. Scale bar equals 500 μm . **B-C.** *Latimeria menadoensis* (MZB 10003). Detail of the odontodes and ridged surface of the superficial layer between the odontodes. Scale bar equals 100 μm . **D.** *Latimeria chalumnae* (MNHN-ZA-AC-2012-26). Detail of the odontodes. The posterior margin of the scale is on the upper right of the image. Scale bar equals 100 μm .

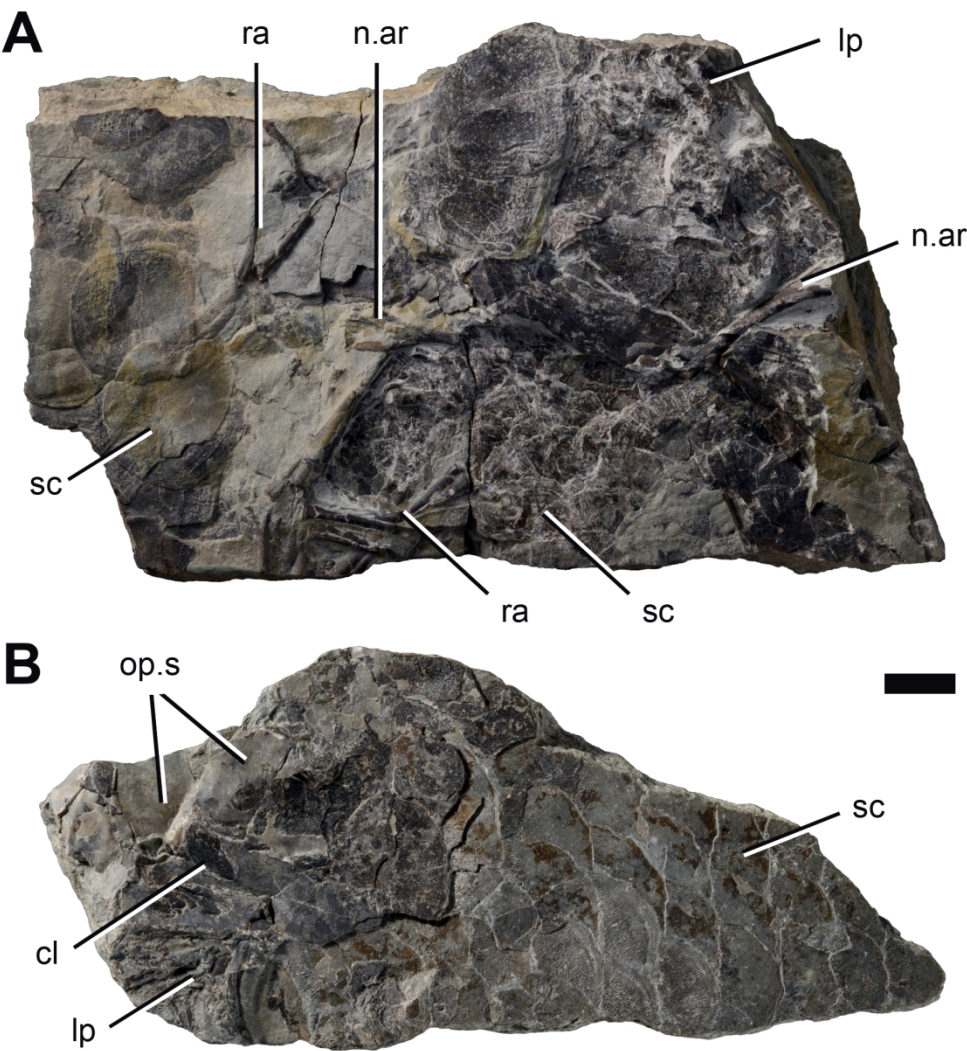
FIGURE 10. Ornamentation and microstructural organization of the scales of *Latimeria*. **A.** *Latimeria chalumnae* (MNHN-ZA-AC-2012-26). Transversal section of the exposed area of the scale in polarized light. The ornamented superficial layer displays eight odontodes (white arrow heads) with large pulp cavities and overlies the basal plate. Scale bar equals 1 mm (from Castanet et al., 1975). **B.** *Latimeria menadoensis* (MZB 10003).

Longitudinal section of the exposed area in microradiography (anterior to the right). The ornamented superficial layer displays four odontodes (white arrowheads). Some vascular canals from the reduced spongiosa can be seen at the base of the odontodes. The whole basal plate is unmineralized and thus does not appear in the image, its approximative contour is represented with a white dashed line. Scale bar equals 1 mm (from Meunier et al., 2008). **Abbreviations:** **bp**, basal plate; **pc**, pulp cavity; **vc**, vascular canal.

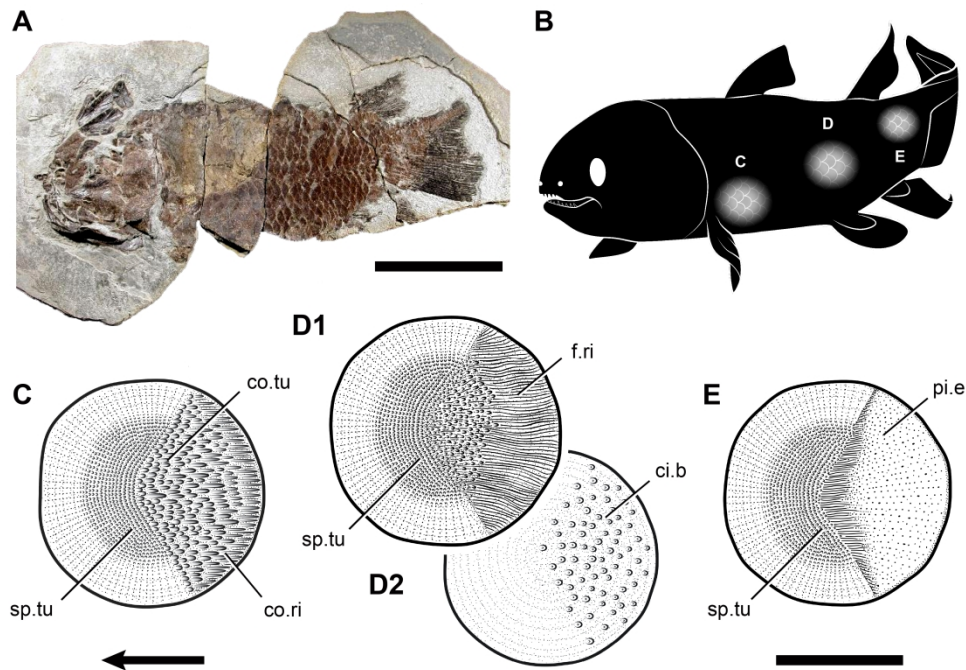
FIGURE 11. Evolution of scale morphology and ornamentation in coelacanth. Phylogenetic hypothesis after Cavin et al. 2017. The date of the diverging nodes is hypothetical. Original artwork of *Miguashaia* and *Latimeria* and modified drawings of *Wimania* after Stensiö, 1921; *Diplocercides* after Stensiö, 1937; *Diplurus* after Schaeffer, 1952; *Rhabdoderma*, *Sassenia*, *Whiteia*, *Lualabaea*, *Axelrodichthys*, *Holophagus*, *Macropoma* after Forey, 1998; *Guizhoucoelacanthus* after Geng et al., 2009. **Nodes:** **1.** Actinistia; **2.** Coelacanthiformes; **3.** Latimeriioidei; **4.** Mawsoniidae; **5.** Latimeriidae. Anterior of the scales to the left.

FIGURE 12. Phylogenetic hypothesis on early actinistian coelacanth evolution with special emphasis on the squamation. **A.** Phylogenetic interrelationships of selected osteichthyans, highlighting the unresolved position of *Styloichthys* and

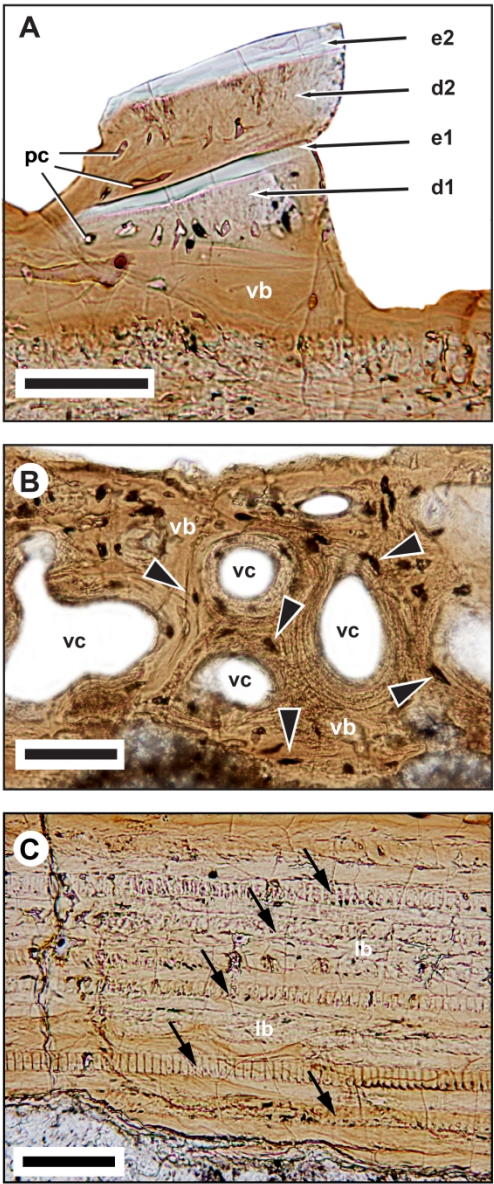
onychodonts (represented here by *Selenodus*) relative to coelacanth (including here *Miguashaia* and *Latimeria*) (consensus of phylogenetic hypotheses after Friedman, 2007; Lu et al., 2016a,b, 2017; Mondéjar-Fernández, 2020, among others). **Abbreviations:** **A:** actinopterygians, **C:** coelacanth, **D:** dipnomorphs, **R:** rhipidistians, **S:** sarcopterygians, **T:** tetrapodomorphs. Original artwork of *Miguashaia*, *Latimeria* and *Danio* and modified drawings of *Andreolepis* after Gross, 1968; *Neoceratodus* after Brien, 1962; *Porolepis*, *Osteolepis* and *Eusthenopteron* after Jarvik, 1980; *Cheirolepis* after Pearson, 1982; *Styloichthys* after Lu & Zhu, 2008; *Psarolepis* after Qu et al., 2013; *Selenodus* after Mondéjar-Fernández, 2020. Not to scale. **B-E.** Phylogenetic hypothesis of the possible interrelationships between *Styloichthys*, onychodonts (irrespective of their monophyly) and coelacanth. **B.** Onychodonts as stem coelacanth and *Styloichthys* as the sister group of coelacanth. **C.** *Styloichthys* as the sister group of the onychodont-coelacanth clade. **D.** *Styloichthys* as the sister group of the onychodont-coelacanth clade and rhipidistians. **E.** Onychodonts as the sister group of coelacanth (including *Styloichthys*) and rhipidistians.



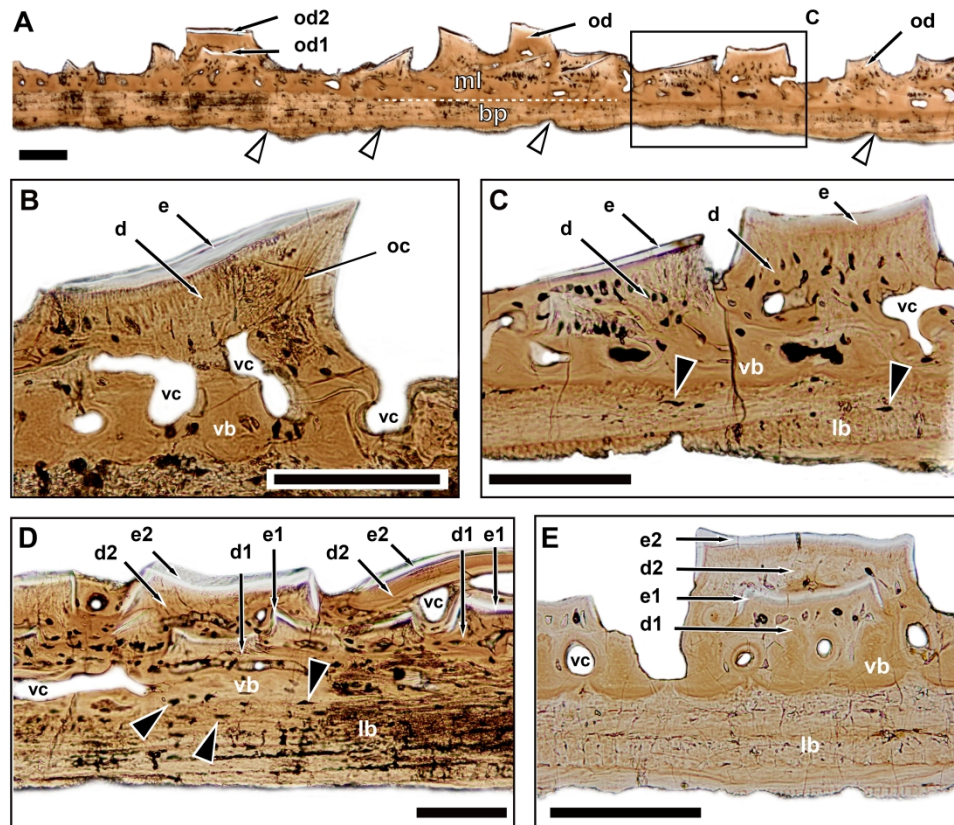
Photographs of the fossil specimens of *Miguashaia bureaui* histologically surveyed. A. MHNM 06-1238. B. MHNM 06-1495. Abbreviations: cl, cleithrum; lp, lepidotrichia; n.ar, neural arches; op.s, opercular series; ra, radials, sc, scales. Scale bar equals 10 mm.



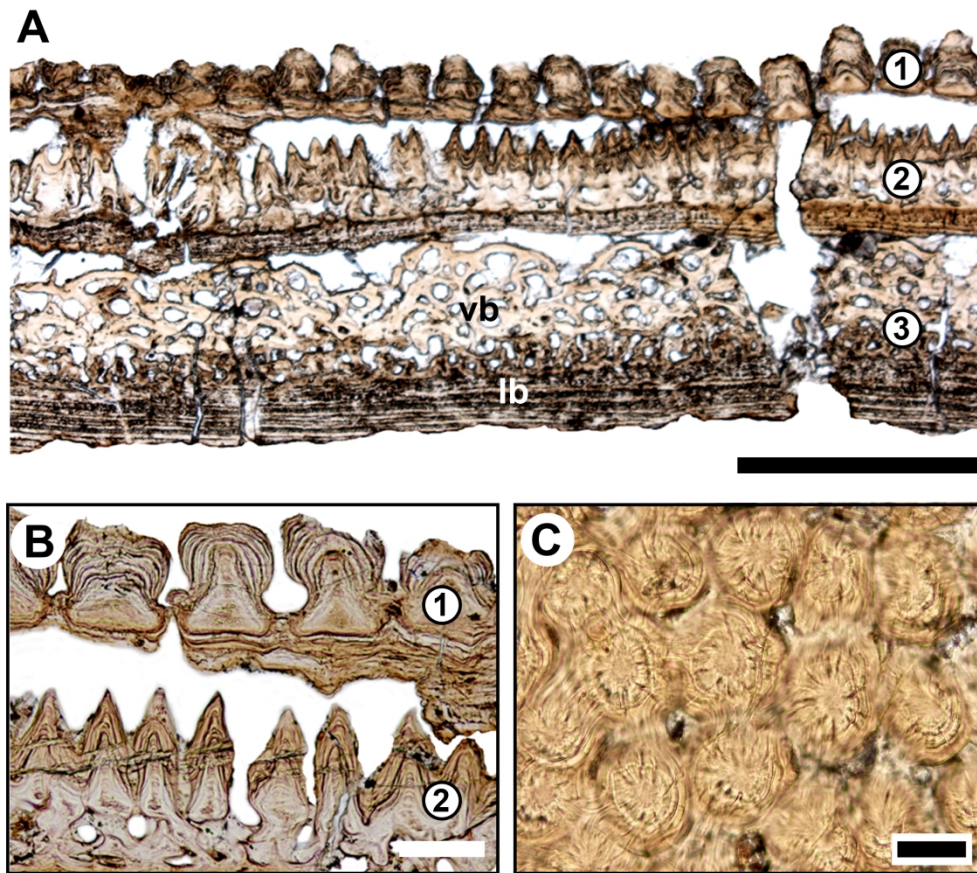
Squamation of *Miguashaia bureauui*. A. Complete specimen (MHNM 06-494). B. Reconstruction and body outline of *M. bureauui* with the scales from three locations of the left flank highlighted. C-E. Reconstruction of different scales from the left flank: C. Scale from the anteroventral portion of the trunk, posterior to the pectoral fin, in external view, D. Scale from the posterior portion of the trunk, in external (1) and internal (2) view. E. Scale from the posterior-most region of the trunk, close to the caudal fin, in external view. Drawings based on MHNM 06-6, MHNM 06-641 (Cloutier 1996; fig. 17), MHNM 06-1238, MHNM 06-1239, and MHNM 06-1495. Abbreviations: ci.b, circular bumps; co.tu, coarse tubercles; co.ri, coarse ridges; f.ri, fine ridges; pi.e, pitted enamel; sp.tu, spoon-shaped tubercles. Arrow points anteriorly. Scale bars equal 10 cm (A) and 10 mm (C-E).



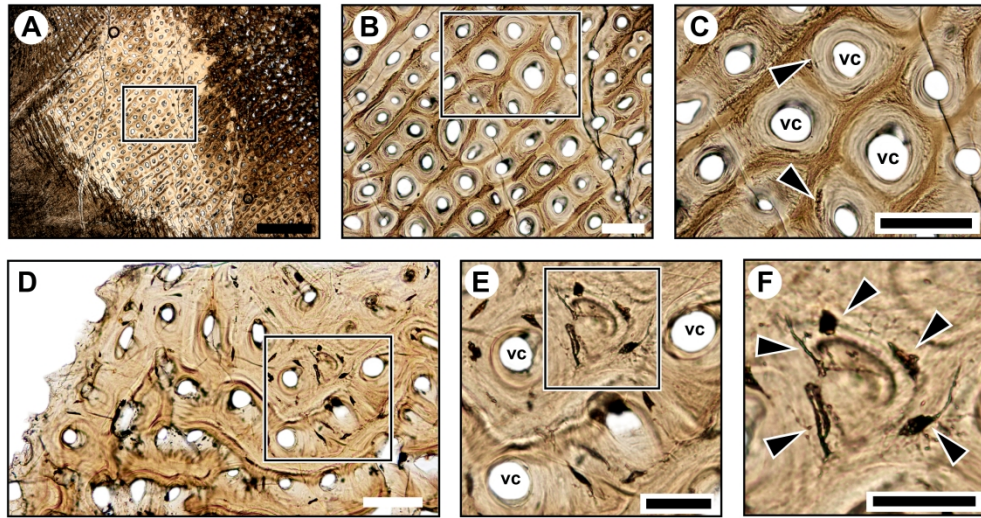
General microstructural organization of the scales of *Miguashaia bureaui* (vertical cross sections of MHNM 06-1495). A. Superimposed odontodes from the superficial layer. B. Vascular bone (spongiosa) from the superficial layer. Arrow heads point to osteocytes embedded in the bone. C. Lamellar bone (isopedine) from the basal plate. Black arrows indicate the repetitive pattern of collagenous plies with the same orientation. Abbreviations: d1-2, dentine layer of two generations of odontodes; e1-2, enamel layer of two generations of odontodes; lb, lamellar bone; pc, pulp cavity; vb, vascular bone. Scale bar equals 50 μm .



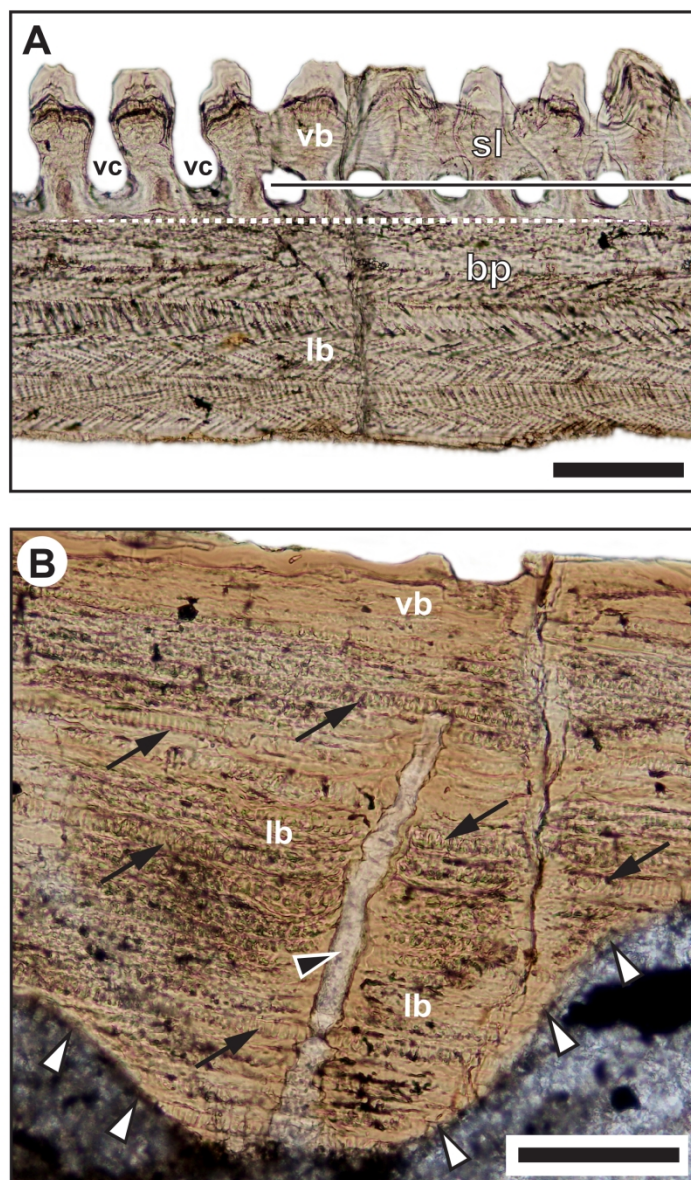
Ornamentation and microstructural organization of the exposed area of the scales of *Miguashaia bureau* (vertical cross sections of MHNM 06-1495). A. General view of a scale showing the distribution of the odontodes in the superficial layer. White arrow heads indicate the irregular front of mineralization of the ventral surface of the basal plate. B. Detail of an odontode composed of a thin layer of enamel and dentine with numerous odontoblastic canaliculi overlying the layer of vascular bone (spongiosa). C. Inset of two odontodes from A. Note the opening of vascular canals from the spongiosa through pores in the surface, as in B, and the large odontoblastic lacunae. Arrow heads indicate the occurrence of osteocyte lacunae in the lamellar bone (isopedine) of the basal plate. D-E. Different distributions and arrangements of two generations of superimposed odontodes. Note the reduced thickness of the spongiosa and the occurrence of vascular canals and vascular bone surrounding the older odontodes. Abbreviations: bp, basal plate; d, dentine; d1-2, dentine layer of two generations of odontodes; e, enamel; e1-2, enamel layer of two generations of odontodes; lb, lamellar bone; ml, middle layer; oc, odontoblastic canaliculi; od1-2, two generations of odontodes; vb, vascular bone; vc, vascular canal; . Scale bars equal 100 μm .



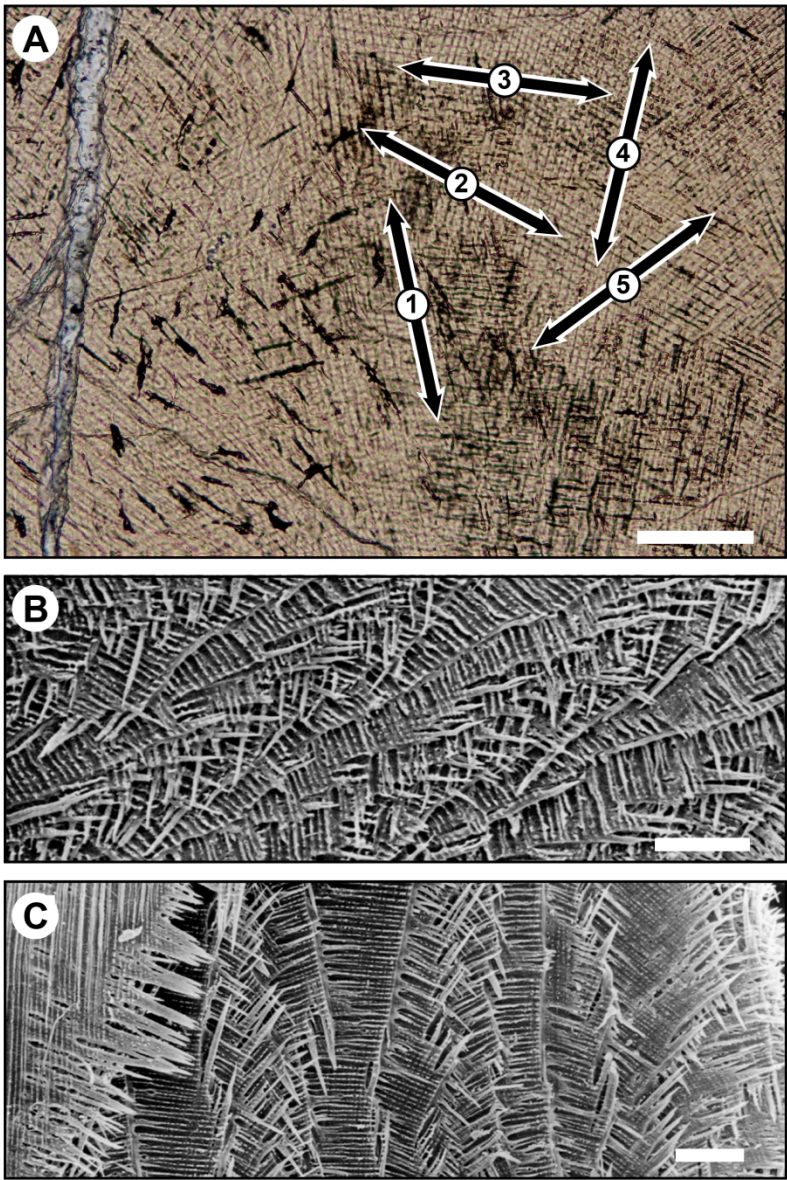
Ornamentation and microstructural organization of the overlapped area of the scales of *Miguashaia bureaui* (vertical and horizontal cross sections of MHNM 06-1238). A. Three scales vertically sectioned at different regions of the overlapped area (from anterior to posterior): 1) Anterior most margin; 2) Central region; 3) Posterior region, close to the contact with the exposed area. Note the increase in thickness of the vascular bone layer (spongiosa) and the lamellar bone layer (isopedine) from anterior to posterior and the changes in the bony ornamentation. Scale bar equals 500 μm . B. Bony ornamentations of the overlapped area from the anterior most margin (1) and the central region (2). Scale bar equals 100 μm . C. Horizontal cross section through the small bony tubercles of the overlapped area from the anterior most margin of a scale (1). Note the concentric patterns of ossification, the radial arrangement of osteocyte lacunae, and the openings of vascular canals between the tubercles. Scale bar equals 50 μm . Abbreviations: lb, lamellar bone; vb, vascular bone.



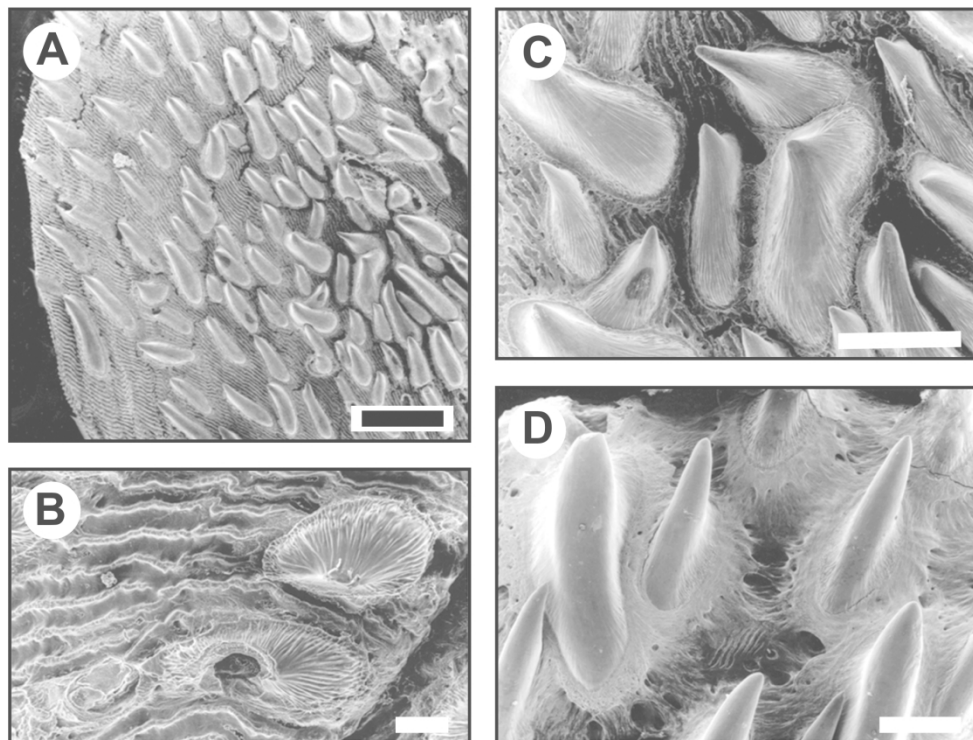
Microstructural and histological organization of the vascular bone of the scales of *Miguashaia bureau* (horizontal cross sections of MHNM 06-1238). A. General view of the vascular bone layer (spongiosa) from the overlapped area of the scales. Note the uniform diameter and distribution of the vascular canals. Scale bar equals 500 μm . B. Detailed inset from A. Note that each vascular canal is surrounded by a series of concentric layers of woven-fibered bone (light brown) while each row of vascular canals is separated from the other by a denser bone tissue (dark brown). Scale bar equals 100 μm . C. Detailed inset from B. Arrow heads indicate the occurrence of osteocyte lacunae surrounding the vascular canals. Scale bar equals 50 μm . D. General view of the vascular bone and vascular canals from another region of the overlapped area of a scale, probably from a more posterior region than in A. Note the more irregular distribution of the vascular canals and the occurrence of ossification waves. Scale bar equals 100 μm . E. Detailed inset from D. Note the occurrence of large osteocyte lacunae in the woven-fibered bone around the vascular canals. Scale bar equals 50 μm . F. Detailed inset from D. Arrow heads point to embedded osteocyte lacunae with large osteocytic prolongations. Scale bar equals 50 μm .



Microstructural and histological organization of the basal plate of the scales of *Miguashaia bureaui* (vertical cross sections of MHNM 06-1238 and MHNM 06-1495). A. General view of the vascular bone layer (spongiosa) of the superficial layer and lamellar bone (isopedine) of the basal plate from the anterior portion of the overlapped area of the scales (MHNM 06-1238). The horizontal plane corresponds to the approximative cross section plane in Fig. 6D-F. Note the clear differences in bone structure between the spongiosa (made of woven-fibered bone) and the isopedine (made of lamellar bone). Scale bar equals 100 μm . B. Detailed view of the basal plate from the central portion of the exposed area of the scales (MHNM 06-1495). Black arrows indicate the repetitive pattern of collagenous plies with the same orientation. White arrow heads indicate the irregular front of mineralization of the visceral surface of the basal plate. The black arrow head points to a vertical vascular canal piercing the basal plate and opening through the ventral surface of the scale. Note that the thickness of the basal plate is more important around the vascular canal resulting in a bump of the internal surface of the scale at this point. Scale bar equals 100 μm . Abbreviations: bp, basal plate; lb, lamellar bone; sl, superficial layer; vb, vascular bone; vc, vascular canal.



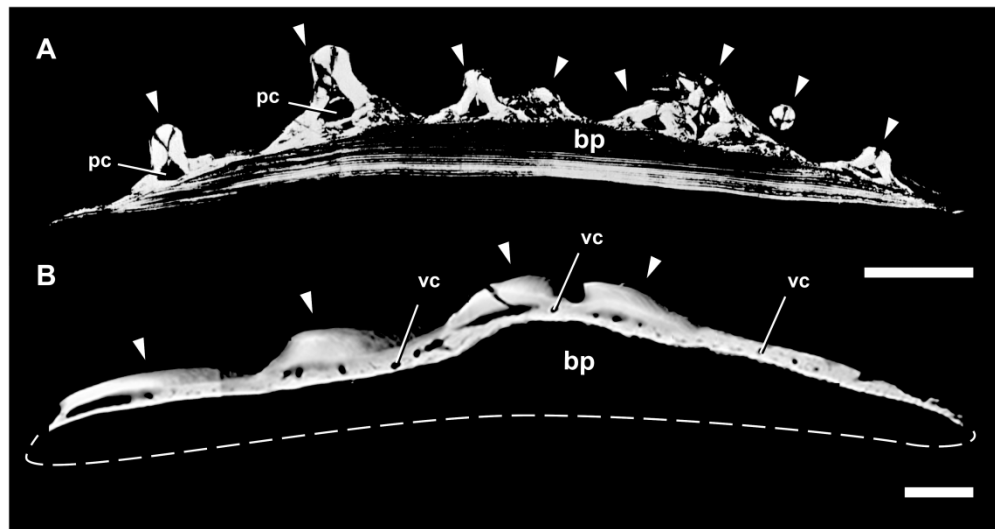
Basal plate comparison of the scales of *Miguashaia* and *Latimeria* in horizontal cross section. A. *Miguashaia* bureaui (MHNM 06-1238). Note the occurrence of 5 directions of the collagen fibers. B. *Latimeria chalumnae* (MNHN-ZA-AC-2012-29). C. *Latimeria menadoensis* (MZB 10003). Note the arciform pattern of the collagen plies evidencing a twisted-plywood arrangement of the layers of the isopedine. Scale bars equal 100 μ m.



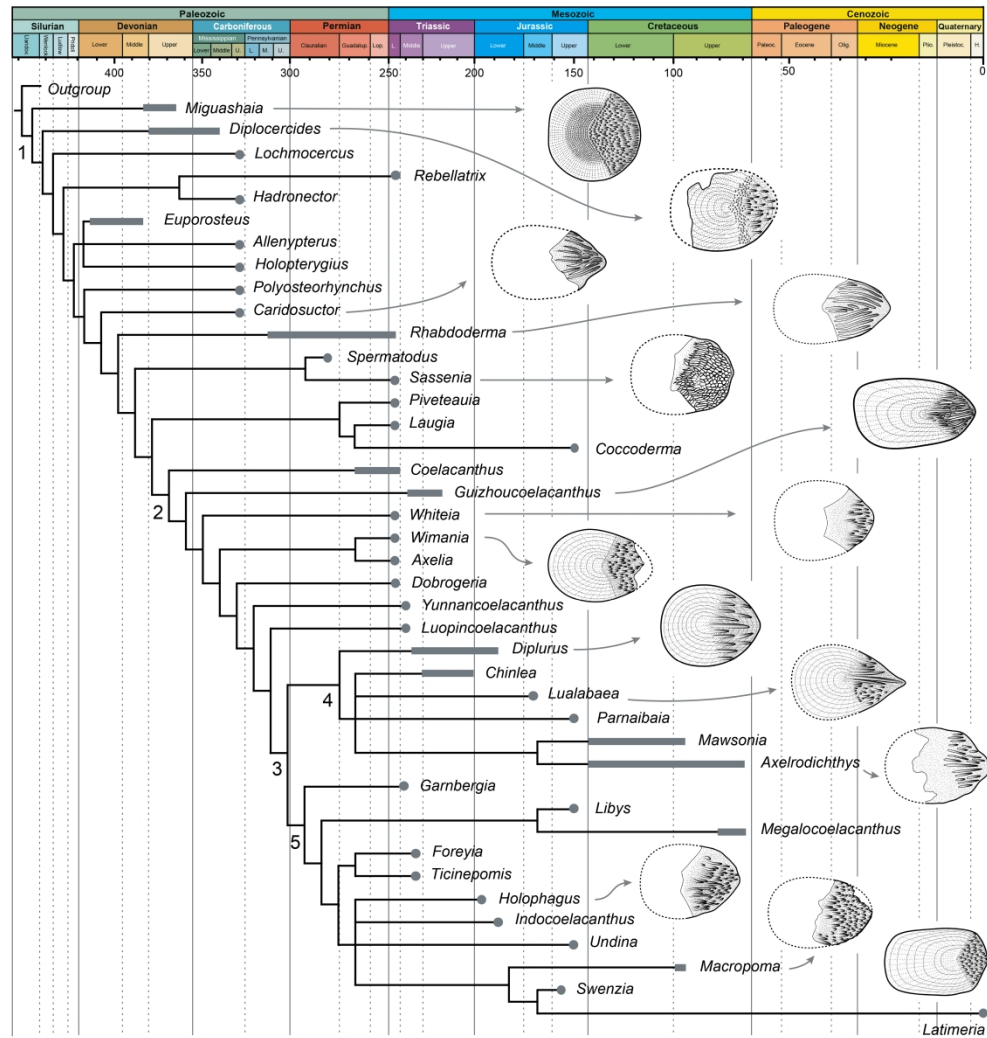
Odontodes from the exposed area of the scales of *Latimeria* (SEM). A. *Latimeria menadoensis* (MZB 10003).

General view of the odontodes. The posterior margin of the scale is on the upper left of the image. The radial ridges of the superficial layer are clearly seen between the odontodes. Scale bar equals 500 μm . B-C.

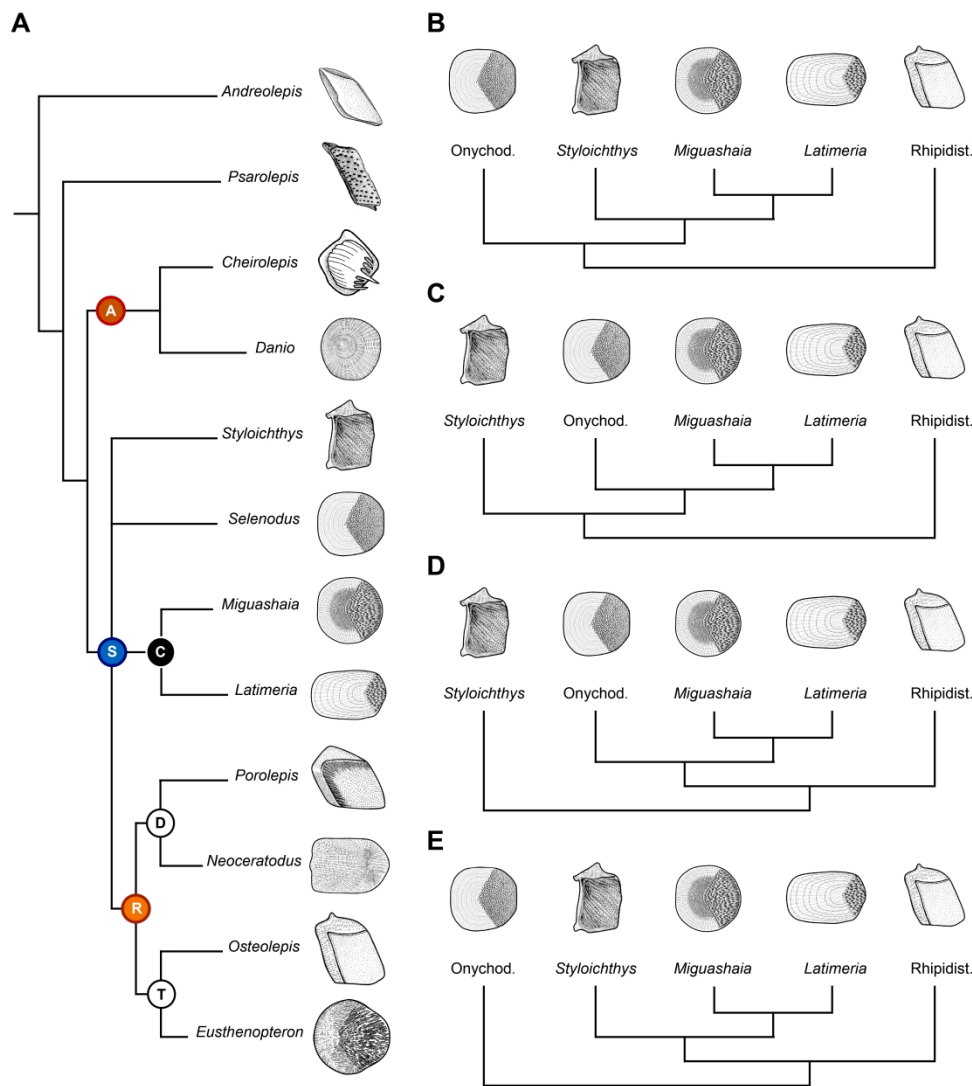
Latimeria menadoensis (MZB 10003). Detail of the odontodes and ridged surface of the superficial layer between the odontodes. Scale bar equals 100 μm . D. *Latimeria chalumnae* (MNHN 1989–806). Detail of the odontodes. The posterior margin of the scale is on the upper right of the image. Scale bar equals 100 μm .



Ornamentation and microstructural organization of the scales of *Latimeria*. A. *Latimeria chalumnae* (MNHN-ZA-AC-2012-26). Transversal section of the exposed area of the scale in polarized light. The ornamented superficial layer displays eight odontodes (white arrow heads) with large pulp cavities and overlies the basal plate. Scale bar equals 1 mm (from Castanet et al. 1975). B. *Latimeria menadoensis* (MZB 10003). Longitudinal section of the exposed area in microradiography (anterior to the right). The ornamented superficial layer displays four odontodes (white arrowheads). Some vascular canals from the reduced spongiosa can be seen at the base of the odontodes. The whole basal plate is unmineralized and thus does not appear in the image, its approximative contour is represented with a white dashed line. Scale bar equals 1 mm (from Meunier et al. 2008). Abbreviations: bp, basal plate; pc, pulp cavity; vc, vascular canal.



Evolution of scale morphology and ornamentation in coelacanths. Phylogenetic hypothesis after Cavin et al. 2017. The date of the diverging nodes is hypothetical. Original artwork of Miguashaia and Latimeria and modified drawings of Wimania after Stensiö 1921; Diplocercides after Stensiö 1937; Diplurus after Schaeffer 1952; Rhabdoderma, Sassenia, Whiteia, Lualabaea, Axelrodichthys, Holophagus, Macropoma after Forey 1998; Guizhoucoelacanthus after Geng et al. 2009. Nodes: 1. Actinistia; 2. Coelacanthiformes; 3. Latimerioidei; 4. Mawsoniidae; 5. Latimeriidae. Anterior of the scales to the left.



Phylogenetic hypothesis on early coelacanth evolution with special emphasis on the squamation. A. Phylogenetic interrelationships of selected osteichthyans, highlighting the unresolved position of Styloichthys and onychodonts (represented here by Selenodus) relative to coelacanths (including here Miguashaia and Latimeria) (consensus of phylogenetic hypotheses after Friedman 2007; Lu et al. 2016a,b, 2017; Mondéjar-Fernández 2020, among others). Abbreviations: A: actinopterygians, C: coelacanths D: dipnomorphs, R: rhipidistians, S: sarcopterygians, T: tetrapodomorphs. Original artwork of Miguashaia, Latimeria and Danio and modified drawings of Andreolepis after Gross 1968; Neoceratodus after Brien, 1962; Porolepis, Osteolepis and Eusthenopteron after Jarvik, 1980; Cheirolepis after Pearson 1982; Styloichthys Lu & Zhu 2008; Psarolepis after Qu et al. 2013; Selenodus after Mondéjar-Fernández 2020. Not to scale. B-E. Phylogenetic hypothesis of the possible interrelationships between Styloichthys, onychodonts (irrespective of their monophyly) and coelacanths. B. Onychodonts as stem coelacanths and Styloichthys as the sister group of coelacanths. C. Styloichthys as the sister group of the onychodont-coelacanth clade. D. Styloichthys as the sister group of the onychodont-coelacanth clade and rhipidistians. E. Onychodonts as the sister group of coelacanths (including Styloichthys) and rhipidistians.

A microanatomical and histological study of the scales of the Devonian sarcopterygian *Miguashaia bureaui* and the evolution of the squamation in coelacanths

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HIGHLIGHTS

- The microstructure of the scales of the early coelacanth *Miguashaia bureaui* from the Devonian of Canada is revealed for the first time.
- The rounded scales display superimposed tubercles of dentine capped with enamel (odontodes) and a fully ossified basal plate.
- *Miguashaia* illustrates the primitive condition for coelacanths and reveals that the general histological arrangement of the coelacanth dermoskeleton has not significantly varied in the last 400 million years, contrary to the scale morphology and ornamentation, which has been more variable.
- Data on the squamation have important implications for the phylogenetic reconstruction of the evolutionary history of coelacanths.